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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification 7:
 C12N 15/12, 15/53, C07K 16/40, 16/18, 14/47, C12N 9/02, A61K 38/17, 38/44,

(11) Interi

(11) International Publication Number:

WO 00/28031

A2 |

(43) International Publication Date:

18 May 2000 (18.05.00)

(21) International Application Number:

PCT/US99/26592

(22) International Filing Date:

48/00, G01N 33/50

10 November 1999 (10.11.99)

(30) Priority Data:

60/107,911 10 November 1998 (10.11.98) US 60/149,332 17 August 1999 (17.08.99) US 60/151,242 27 August 1999 (27.08.99) US

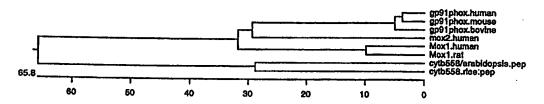
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(81) Designated States: AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK, DM, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, UZ, VN, YU, ZA, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

Published

Without international search report and to be republished upon receipt of that report.

(54) Title: NOVEL MITOGENIC REGULATORS



(57) Abstract

The present invention relates to new genes encoding for the production of novel proteins involved in generation of reactive oxygen intermediates that affect cell division. The present invention also provides vectors containing these genes, cells transfected with these vectors, antibodies raised against these novel proteins, kits for detection, localization and measurement of these genes and proteins, and methods to determine the activity of drugs to affect the activity of the proteins of the present invention.

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NOVEL MITOGENIC REGULATORS

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TECHNICAL FIELD

The present invention relates to the field of normal and abnormal cell growth, in particular mitogenic regulation. The present invention provides the following: nucleotide sequences encoding for the production of enzymes that are mitogenic regulators; amino acid sequences of these enzymes; vectors containing these nucleotide sequences; methods for transfecting cells with vectors that produce these enzymes; transfected cells; methods for administering these transfected cells to animals to induce tumor formation; and antibodies to these enzymes that are useful for detecting and measuring levels of these enzymes, and for binding to cells possessing extracellular epitopes of these enzymes.

BACKGROUND OF THE INVENTION

Reactive oxygen intermediates (ROI) are partial reduction products of oxygen: 1 electron reduces O_2 to form superoxide (O_2) , and 2 electrons reduce O_2 to form hydrogen peroxide (H_2O_2) . ROI are generated as a byproduct of aerobic metabolism and by toxicological mechanisms. There is growing evidence for regulated enzymatic generation of O_2 and its conversion to H_2O_2 in a variety of cells. The conversion of O_2 to H_2O_2 occurs spontaneously, but is markedly accelerated by superoxide dismutase (SOD). High levels of ROI are associated with damage to biomolecules such as DNA, biomembranes and proteins. Recent evidence indicates generation of ROI under

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normal cellular conditions and points to signaling roles for O_2 and H_2O_2 .

Several biological systems generate reactive oxygen. Phagocytic cells such as neutrophils generate large quantities of ROI as part of their battery of bactericidal mechanisms. Exposure of neutrophils to bacteria or to various soluble mediators such as formyl-Met-Leu-Phe or phorbol esters activates a massive consumption of oxygen, termed the respiratory burst, to initially generate superoxide, with secondary generation of H₂O₂, HOCl and hydroxyl radical. The enzyme responsible for this oxygen consumption is the respiratory burst oxidase (nicotinamide adenine dinucleotide phosphate-reduced form (NADPH) oxidase).

There is growing evidence for the generation of ROI by non-phagocytic cells, particularly in situations related to cell proliferation. Significant generation of H₂O₂, O₂, or both have been noted in some cell types. Fibroblasts and human endothelial cells show increased release of superoxide in response to cytokines such as interleukin-1 or tumor necrosis factor (TNF) (Meier et al. (1989) Biochem J. 263, 539-545.; Matsubara et al. (1986) J. Immun. 137, 3295-3298). transformed fibroblasts show increased superoxide release compared with control fibroblasts (Irani, et al. (1997) Science 275, 1649-1652). Rat vascular smooth muscle cells show increased H₂O₂ release in response to PDGF (Sundaresan et al. (1995) Science 270, 296-299) and angiotensin II (Griendling et al. (1994) Circ. Res. 74, 1141-1148; Fukui et al. (1997) Circ. Res. 80, 45-51; Ushio-Fukai et al. (1996) J. Biol. Chem. 271, 23317-23321), and H₂O₂ in these cells is associated with increased proliferation rate. The occurrence of ROI in a variety of cell types is summarized in Table 1 (adapted from Burdon, R. (1995) Free Radical Biol. Med. 18, 775-794).

Table 1

Superoxide
human fibroblasts
human endothelial cells
human/rat smooth muscle cells
human fat cells
human osteocytes
human osteocytes
human colonic epithelial cells

Hydrogen Peroxide
Balb/3T3 cells
rat pancreatic islet cells
murine keratinocytes
rabbit chondrocytes
human tumor cells
fat cells, 3T3 L1 cells

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ROI generated by the neutrophil have a cytotoxic function. While ROI are normally directed at the invading microbe, ROI can also induce tissue damage (e.g., in inflammatory conditions such as arthritis, shock, lung disease, and inflammatory bowel disease) or may be involved in tumor initiation or promotion, due to damaging effects on DNA. Nathan (Szatrowski et al. (1991) Canc. Res. 51, 794-798) proposed that the generation of ROI in tumor cells may contribute to the hypermutability seen in tumors, and may therefore contribute to tumor heterogeneity, invasion and metastasis.

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In addition to cytotoxic and mutagenic roles, ROI have ideal properties as signal molecules: 1) they are generated in a controlled manner in response to upstream signals; 2) the signal can be terminated by rapid metabolism of O_2 and H_2O_2 by SOD and catalase/peroxidases; 3) they elicit downstream effects on target molecules, e.g., redox-sensitive regulatory proteins such as NF kappa B and AP-1 (Schreck et al. (1991) *EMBO J.* 10, 2247-2258; Schmidt et al. (1995) *Chemistry & Biology* 2, 13-22). Oxidants such as O_2 and H_2O_2 have a relatively well defined signaling role in bacteria, operating via the SoxI/II regulon to regulate transcription.

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ROI appear to have a direct role in regulating cell division, and may function as mitogenic signals in pathological conditions related to growth. These conditions include cancer and cardiovascular disease. O_2^- is generated in endothelial cells

in response to cytokines, and might play a role in angiogenesis (Matsubara et al. (1986) J. Immun. 137, 3295-3298). O₂ and H₂O₂ are also proposed to function as "life-signals", preventing cells from undergoing apoptosis (Matsubara et al. (1986) J. Immun. 137, 3295-3298). As discussed above, many cells respond to growth factors (e.g., platelet derived growth factor (PDGF), epidermal derived growth factor (EGF), angiotensin II, and various cytokines) with both increased production of O₂ /H₂O₂ and increased proliferation. Inhibition of ROI generation prevents the mitogenic response. Exposure to exogenously generated O₂ and H₂O₂ results in an increase in cell proliferation. A partial list of responsive cell types is shown below in Table 2 (adapted from Burdon, R. (1995) Free Radical Biol. Med. 18, 775-794).

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Table 2

Superoxide	Hydrogen peroxide
human, hamster fibroblasts	mouse osteoblastic cells
Balb/3T3 cells	Balb/3T3 cells
human histiocytic leukemia	rat, hamster fibroblasts
mouse epidermal cells	human smooth muscle cells
rat colonic epithelial cells	rat vascular smooth muscle cells
rat vascular smooth muscle cells	

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While non-transformed cells can respond to growth factors and cytokines with the production of ROI, tumor cells appear to produce ROI in an uncontrolled manner. A series of human tumor cells produced large amounts of hydrogen peroxide compared with non-tumor cells (Szatrowski et al. (1991) Canc. Res. 51, 794-798). Ras-transformed NIH 3T3 cells generated elevated amounts of superoxide, and inhibition of superoxide generation by several mechanisms resulted in a reversion to a "normal" growth phenotype.

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O₂ has been implicated in maintenance of the transformed phenotype in cancer cells including melanoma, breast carcinoma, fibrosarcoma, and virally transformed tumor

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cells. Decreased levels of the manganese form of SOD (MnSOD) have been measured in cancer cells and in vitro-transformed cell lines, predicting increased O₂ levels (Burdon, R. (1995) Free Radical Biol. Med. 18, 775-794). MnSOD is encoded on chromosome 6q25 which is very often lost in melanoma. Overexpression of MnSOD in melanoma and other cancer cells (Church et al. (1993) Proc. of Natl. Acad. Sci. 90, 3113-3117; Fernandez-Pol et al. (1982) Canc. Res. 42, 609-617; Yan et al. (1996) Canc. Res. 56, 2864-2871) resulted in suppression of the transformed phenotype.

ROI are implicated in growth of vascular smooth muscle associated with hypertension, atherosclerosis, and restenosis after angioplasty. O2 generation is seen in rabbit aortic adventitia (Pagano et al. (1997) Proc. Natl. Acad. Sci. 94, 14483-14488). Vascular endothelial cells release O₂ in response to cytokines (Matsubara et al. (1986) J. Immun. 137, 3295-3298). O₂ is generated by aortic smooth muscle cells in culture, and increased O2 generation is stimulated by angiotensin II which also induces cell hypertrophy. In a rat model system, infusion of angiotensin II leads to hypertension as well as increased O2 generation in subsequently isolated aortic tissue (Ushio-Fukai et al. (1996) J. Biol. Chem. 271, 23317-23321.; Yu et al. (1997) J. Biol. Chem. 272, 27288-27294). Intravenous infusion of a form of SOD that localizes to the vasculature or an infusion of an O2 scavenger prevented angiotensin II induced hypertension and inhibited ROI generation (Fukui et al. (1997) Circ. Res. 80, 45-51).

The neutrophil NADPH oxidase, also known as phagocyte respiratory burst oxidase, provides a paradigm for the study of the specialized enzymatic ROI-generating system. This extensively studied enzyme oxidizes NADPH and reduces oxygen to form O_2 . NADPH oxidase consists of multiple proteins and is regulated by assembly of cytosolic and membrane components. The catalytic moiety consists of flavocytochrome b_{558} , an integral plasma membrane enzyme comprised of two components: gp91phox (gp refers to

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glycoprotein; phox is an abbreviation of the words phagocyte and oxidase) and p22phox (p refers to protein). contains 1 flavin adenine dinucleotide (FAD) and 2 hemes as well as the NADPH binding site. p22phox has a C-terminal proline-rich sequence which serves as a binding site for cytosolic regulatory proteins. The two cytochrome subunits, gp91phox and p22phox appear to stabilize one another, since the genetic absence of either subunit, as in the inherited disorder chronic granulomatous disease (CGD), results in the absence of the partner subunit (Yu et al. (1997) J. Biol. Chem. 272, 27288-27294). Essential cytosolic proteins include p47phox, p67phox and the small GTPase Rac, of which there are two isoforms. p47phox and p67phox both contain SH₃ regions and proline-rich regions which participate in protein interactions assembly of the oxidase components during governing activation. The neutrophil enzyme is regulated in response to bacterial phagocytosis or chemotactic signals phosphorylation of p47phox, and perhaps other components, as well as by guanine nucleotide exchange to activate the GTPbinding protein Rac.

The origin of ROI in non-phagocytic tissues is unproven, but the occurrence of phagocyte oxidase components has been evaluated in several systems by immunochemical methods, Northern blots and reverse transcriptase-polymerase chain reaction (RT-PCR). The message for p22phox is expressed widely, as is that for Rac1. Several cell types that are capable of O₂ generation have been demonstrated to contain all of the phox components including gp91phox, as summarized below in Table 3. These cell types include endothelial cells, aortic adventitia and lymphocytes.

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Table 3

Tissue	gp91phox	p22phox	p47phox	p67phox
neutrophil	+1.2	+1,2	+1,2	+1.2
aortic adventitia	+1	+1	+1	+1
lymphocytes	+2	+2	+1.2	+1.2
endothelial cells	+2	+ ²	+1,2	+1,2
glomerular mesangial cells	-	+1.2	+1,2	+1,2
fibroblasts	-	+2	+1.2	+2
aortic sm. muscle	-	+1.2	?	?

l= protein expression shown. 2= mRNA expression shown.

However, a distinctly different pattern is seen in several other cell types shown in Table 3 including glomerular mesangial cells, rat aortic smooth muscle and fibroblasts. these cells, expression of gp91phox is absent while p22phox and in some cases cytosolic phox components have been demonstrated to be present. Since gp91phox and p22phox stabilize one another in the neutrophil, there has been much speculation that some molecule, possibly related to gp91phox, accounts for ROI generation in glomerular mesangial cells, rat aortic smooth muscle and fibroblasts (Ushio-Fukai et al. (1996) J. Biol. Chem. 271, 23317-23321). Investigation of fibroblasts from a patient with a genetic absence of gp91phox provides proof that the gp91phox subunit is not involved in ROI generation in these cells (Emmendorffer et al. (1993) Eur. J. Haematol. 51, 223-227). Depletion of p22phox from vascular smooth muscle using an antisense approach indicated that this subunit participates in ROI generation in these cells, despite the absence of detectable gp91phox (Ushio-Fukai et al. (1996) J. Biol. Chem. 271, 23317-23321). At this time the molecular candidates possibly related to gp91phox and involved in ROI generation in these cells are unknown.

Accordingly, what is needed is the identity of the proteins involved in ROI generation, especially in non-phagocytic tissues and cells. What is also needed are the

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nucleotide sequences encoding for these proteins, and the primary sequences of the proteins themselves. Also needed are vectors designed to include nucleotides encoding for these proteins. Probes and PCR primers derived from the nucleotide sequence are needed to detect, localize and measure nucleotide sequences, including mRNA, involved in the synthesis of these proteins. In addition, what is needed is a means to transfect cells with these vectors. What is also needed are expression systems for production of these molecules. Also needed are antibodies directed against these molecules for a variety of uses including localization, detection, measurement and passive immunization.

SUMMARY OF THE INVENTION

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The present invention solves the problems described above by providing a novel family of nucleotide sequences and proteins encoded by these nucleotide sequences termed mox proteins and duox proteins. In particular the present invention provides compositions comprising nucleotide sequences SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:22, SEQ ID NO:41, SEQ ID NO:45, SEQ ID NO:47, and fragments thereof, which encode for the expression of proteins comprising SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:21, SEQ ID NO:42, SEQ ID NO:46, SEQ ID NO:48, respectively, and fragments thereof. While not wanting to be bound by the following statement, it is believed that these proteins are involved in ROI production. The present invention also provides vectors containing these nucleotide sequences, cells transfected with these vectors which produce the proteins comprising SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:21, SEQ ID NO:42, SEQ ID NO:46, SEQ ID NO:48, and fragments thereof, and antibodies to these proteins and fragments thereof. The present invention also provides methods for stimulating cellular proliferation by administering vectors encoded for production of the proteins comprising SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:21, SEQ ID NO:42, SEQ ID NO:46, SEQ ID NO:48 and fragments thereof. The present invention also

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provides methods for stimulating cellular proliferation by administering the proteins comprising SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:45, SEQ ID NO:46, SEQ ID NO:48 and fragments thereof. The nucleotides and antibodies of the present invention are useful for the detection, localization and measurement of the nucleic acids encoding for the production of the proteins of the present invention, and also for the detection, localization and measurement of the proteins of the present invention. These nucleotides and antibodies can be combined with other reagents in kits for the purposes of detection, localization and measurement.

Most particularly, the present invention involves a method for regulation of cell division or cell proliferation by modifying the activity or expression of the proteins described as SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:21, SEQ ID NO:42, SEQ ID NO:46, SEQ ID NO:48 or fragments thereof. These proteins, in their naturally occurring or expressed forms, are expected to be useful in drug development, for example for screening of chemical and drug libraries by observing inhibition of the activity of these enzymes. Such chemicals and drugs would likely be useful as treatments for cancer, prostatic hypertrophy, benign prostatic hypertrophy, hypertension, atherosclerosis and many other disorders involving abnormal cell growth or proliferation as described below. The entire expressed protein may be useful in these assays. Portions of the molecule which may be targets for inhibition or modification include but are not limited to the binding site for pyridine nucleotides (NADPH or NADH), the flavoprotein domain (approximately the C-terminal 265 amino acids), and/or the binding or catalytic site for flavin adenine dinucleotide (FAD).

The method of the present invention may be used for the development of drugs or other therapies for the treatment of conditions associated with abnormal growth including, but not limited to the following: cancer, psoriasis, prostatic hypertrophy, benign prostatic hypertrophy, cardiovascular disease, proliferation of vessels, including but

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not limited to blood vessels and lymphatic vessels, arteriovenous malformation, vascular problems associated with the eye, atherosclerosis, hypertension, and restenosis following angioplasty. The enzymes of the present invention are excellent targets for the development of drugs and other agents which may modulate the activity of these enzymes. It is to be understood that modulation of activity may result in enhanced, diminished or absence of enzymatic activity. Modulation of the activity of these enzymes may be useful in treatment of conditions associated with abnormal growth.

Drugs which affect the activity of the enzymes represented in SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:21, SEQ ID NO:42, SEQ ID NO:46, SEQ ID NO:48, or fragments thereof, may also be combined with other therapeutics in the treatment of specific conditions. For example, these drugs may be combined with angiogenesis inhibitors in the treatment of cancer, with antihypertensives for the treatment of hypertension, and with cholesterol lowering drugs for the treatment of atherosclerosis.

Accordingly, an object of the present invention is to provide nucleotide sequences, or fragments thereof, encoding for the production of proteins, or fragments thereof, that are involved in ROI production.

Another object of the present invention is to provide vectors containing these nucleotide sequences, or fragments thereof.

Yet another object of the present invention is to provide cells transfected with these vectors.

Still another object of the present invention is to administer cells transfected with these vectors to animals and humans.

Another object of the present invention is to provide proteins, or fragments thereof, that are involved in ROI production.

Still another object of the present invention is to provide antibodies, including monoclonal and polyclonal

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antibodies, or fragments thereof, raised against proteins, or fragments thereof, that are involved in ROI production.

Another object of the present invention is to administer genes containing nucleotide sequences, or fragments thereof, encoding for the production of proteins, or fragments thereof, that are involved in ROI production, to animals and humans and also to cells obtained from animals and humans.

Another object of the present invention is to administer antisense complimentary sequences of genes containing nucleotide sequences, or fragments thereof, encoding for the production of proteins, or fragments thereof, that are involved in ROI production, to animals and humans and also to cells obtained from animals and humans.

Yet another object of the present invention is to provide a method for stimulating or inhibiting cellular proliferation by administering vectors containing nucleotide sequences, or fragments thereof, encoding for the production of proteins, or fragments thereof, that are involved in ROI production, to animals and humans. It is also an object of the present invention to provide a method for stimulating or inhibiting cellular proliferation by administering vectors containing antisense complimentary sequences of nucleotide sequences, or fragments thereof, encoding for the production of proteins, or fragments thereof, that are involved in ROI production, to animals and humans. These methods of stimulating cellular proliferation are useful for a variety of purposes, including but not limited to, developing animal models of tumor formation, stimulating cellular proliferation of blood marrow cells following chemotherapy or radiation, or in cases of anemia.

Still another object of the present invention is to provide antibodies useful in immunotherapy against cancers expressing the proteins represented in SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:45, SEQ ID NO:46, SEQ ID NO:48 or fragments thereof.

Yet another object of the present invention is to provide nucleotide probes useful for the detection, localization and measurement of nucleotide sequences, or fragments thereof, encoding for the production of proteins, or fragments thereof, that are involved in ROI production.

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Another object of the present invention is to provide antibodies useful for the detection, localization and measurement of nucleotide sequences, or fragments thereof, encoding for the production of proteins, or fragments thereof, that are involved in ROI production.

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Another object of the present invention is to provide kits useful for detection of nucleic acids including the nucleic acids represented in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:22, SEQ ID NO:41, SEQ ID NO:45, SEQ ID NO:47, or fragments thereof, that encode for proteins, or fragments thereof, that are involved in ROI production.

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Yet another object of the present invention is to provide kits useful for detection and measurement of nucleic acids including the nucleic acids represented in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:22, SEQ ID NO:41, SEQ ID NO:45, SEQ ID NO:47, or fragments thereof, that encode for proteins, or fragments thereof, that are involved in ROI production.

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Still another object of the present invention is to provide kits useful for the localization of nucleic acids including the nucleic acids represented in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:22, SEQ ID NO:41, SEQ ID NO:45, SEQ ID NO:47, or fragments thereof, that encode for proteins, or fragments thereof that are involved in ROI production.

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Another object of the present invention is to provide kits useful for detection of proteins, including the proteins represented in SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:41, SEQ ID NO:42, SEQ ID NO:46, SEQ ID NO:48, or fragments thereof, that are involved in ROI production.

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Yet another object of the present invention is to provide kits useful for detection and measurement of proteins,

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including the proteins represented in SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:41, SEQ ID NO:42, SEQ ID NO:46, SEQ ID NO:48, or fragments thereof, that are involved in ROI production.

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Still another object of the present invention is to provide kits useful for localization of proteins, including the proteins represented in SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:41, SEQ ID NO:42, SEQ ID NO:46, SEQ ID NO:48, or fragments thereof, that are involved in ROI production.

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Yet another object of the present invention is to provides kits useful for the detection, measurement or localization of nucleic acids, or fragments thereof, encoding for proteins, or fragments thereof, that are involved in ROI production, for use in diagnosis and prognosis of abnormal cellular proliferation related to ROI production.

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Another object of the present invention is to provides kits useful for the detection, measurement or localization of proteins, or fragments thereof, that are involved in ROI production, for use in diagnosis and prognosis of abnormal cellular proliferation related to ROI production.

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These and other objects, features and advantages of the present invention will become apparent after a review of the following detailed description of the disclosed embodiments and the appended drawings.

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BRIEF DESCRIPTION OF THE FIGURES

Fig. 1(a-d). Comparison of amino acid sequences of the human mox1 protein (labeled mox1.human, SEQ ID NO:2), rat mox1 protein (labeled mox1.rat, SEQ ID NO:21), human mox2 protein (labeled mox2.human., SEQ ID NO:4) of the present invention to human (gp 91phox/human.pep, SEQ ID NO:12) bovine (gp 91 phox/bovine.pep, SEQ ID NO:37), and murine (gp 91 phox/mouse.pep, SEQ ID NO:38) proteins. Also included are related plant enzyme proteins cytb 558.arabidopsis.pep (SEQ ID NO:39) and cytb558.rice.pep,

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(SEQ ID NO:40). Enclosed in boxes are similar amino acid residues.

- Fig. 2. Sequence similarities among proteins related to gp91phox including human mox1 (SEQ ID NO:2), human mox2 (SEQ ID NO:4), and rat mox1 (SEQ ID NO:21). The dendrogram indicates the degree of similarity among this family of proteins, and also includes the related plant enzymes.
- Fig. 3. Cell free assay for mox-1 activity. Superoxide generation was measured using the chemiluminescent reaction between lucigenin and superoxide in cell lysates from vector control NEF2 and mox1 transfected NIH3T3 cells.
- Fig. 4. Superoxide generation by human mox1. Reduction of NBT in mox1 transfected and control fibroblasts was measured in the absence (filled bars) or presence (open bars) or superoxide dismutase.
- Fig. 5. Aconitase (filled bars), lactate dehydrogenase (narrow hatching) and fumarase (broad hatching) were determined in lysates of cells transfected with vector alone (NEF2) or with mox1 (YA26, YA28 and YA212).

DETAILED DESCRIPTION OF THE INVENTION

The present invention solves the problems described above by providing a novel family of nucleotide sequences and proteins, encoded by these nucleotide sequences, termed mox proteins and duox proteins. The term "mox" refers to "mitogenic oxidase" while the term "duox" refers to "dual oxidase". In particular, the present invention provides novel compositions comprising the nucleotide sequences SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:22, SEQ ID NO:41, SEQ ID NO:45, SEQ ID NO:47, and fragments thereof, which encode, respectively, for the expression of proteins comprising SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:21, SEQ ID NO:42, SEQ ID NO:46, SEQ ID NO:48 and fragments thereof.

Both the mox and duox proteins described herein have homology to the gp91phox protein involved in ROI

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generation, however, the mox and duox proteins comprise a novel and distinct family of proteins. The mox proteins included in the present invention have a molecular weight of approximately 65 kDa as determined by reducing gel electrophoresis and are capable of inducing ROI generation in cells. As described in more detail below, the mox proteins of the present invention also function in the regulation of cell growth, and are therefore implicated in diseases involving abnormal cell growth such as cancer. The present invention describes mox proteins found in human and rat, however, it is likely that the mox family of genes/proteins is widely distributed among multicellular organisms.

The duox proteins described herein are larger than the mox proteins and have three distinct regions: the amino terminal region having homology to peroxidase proteins, the internal region having homology to calmodulin (CAM) proteins and the carboxy-terminal region having homology to mox proteins. Human duox1 is shown in SEQ ID NO:46 and a portion of human duox2 is shown in SEQ ID NO:48. Nucleotides encoding duox1 and duox2 proteins are also shown in SEQ ID NO: 45 and SEQ ID NO:47, respectively. addition to the human duox proteins, comparison of the sequence of human duox1 and human duox2 with genomic databases using BLAST searching resulted in the identification of two homologs of duox in C. elegans (Ce-duox1 and Ceduox2). Drosophila also appears to have at least one duox homolog. Thus, the duox family of genes/proteins is widely distributed among multicellular organisms.

Although not wanting to be bound by the following statement, it is believed that duox1 and duox2 have dual enzymatic functions, catalyzing both the generation of superoxide and peroxidative type reactions. The latter class of reactions utilize hydrogen peroxide as a substrate (and in some cases have been proposed to utilize superoxide as a substrate). Since hydrogen peroxide is generated spontaneously from the dismutation of superoxide, it is believed that the NAD(P)H

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oxidase domain generates the superoxide and/or hydrogen peroxide which can then be used as a substrate for the peroxidase domain. In support of this hypothesis, a model for the duox1 protein in *C. elegans* has been developed that has an extracellular N-terminal peroxidase domain, a transmembrane region and a NADPH binding site located on the cytosolic face of the plasma membrane. By analogy with the neutrophil NADPH-oxidase which generates extracellular superoxide, human duox1 is predicted to generate superoxide and its byproduct hydrogen peroxide extracellularly where it can be utilized by the peroxidase domain.

While the ROI generated by duox1 and duox2 may function as does mox1 in regulation of cell growth, the presence of the peroxidase domain is likely to confer additional biological functions. Depending upon the co-substrate, peroxidases can participate in a variety of reactions including halogenation such as the generation of hypochlorous acid (HOCl) by myeloperoxidase and the iodination of tyrosine to form thyroxin by thyroid peroxidase. Peroxidases have also been documented to participate in the metabolism of polyunsaturated fatty acids, and in the chemical modification of tyrosine in collagen (by sea urchin ovoperoxidase). Although not wanting to be bound by this statement, it is believed that the predicted transmembrane nature of duox1 facilitates its function in the formation or modification of extracellular matrix or basement membrane. Since the extracellular matrix plays an important role in tumor cell growth, invasion and metastasis, it is believed that the duox type enzymes play a pathogenic role in such conditions.

In addition to the nucleotide sequences described above, the present invention also provides vectors containing these nucleotide sequences and fragments thereof, cells transfected with these vectors which produce the proteins comprising SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:21, SEQ ID NO:42, SEQ ID NO:46, SEQ ID NO:48 and fragments thereof, and antibodies to these proteins and fragments thereof.

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The present invention also provides methods for stimulating cellular proliferation by administering vectors, or cells containing vectors, encoded for production of the proteins comprising SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:21, SEQ ID NO:42, SEQ ID NO:46, SEQ ID NO:48 and fragments thereof. The nucleotides and antibodies of the present invention are useful for the detection, localization and measurement of the nucleic acids encoding for the production of the proteins of the present invention, and also for the detection, localization and measurement of the proteins of the present invention. These nucleotides and antibodies can be combined with other reagents in kits for the purposes of detection, localization and measurement. These kits are useful for diagnosis and prognosis of conditions involving cellular proliferation associated with production of reactive oxygen intermediates.

The present invention solves the problems described above by providing a composition comprising the nucleotide sequence SEQ ID NO:1 and fragments thereof. The present invention also provides a composition comprising the nucleotide sequence SEQ ID NO:3 and fragments thereof. The present invention also provides a composition comprising the nucleotide sequence SEQ ID NO:22 and fragments thereof. The present invention also provides a composition comprising the nucleotide sequence SEQ ID NO:41 and fragments thereof. The present invention also provides a composition comprising the nucleotide sequence SEQ ID NO:45 and fragments thereof. The present invention also provides a composition comprising the nucleotide sequence SEQ ID NO:45 and fragments thereof. The present invention also provides a composition comprising the nucleotide sequence SEQ ID NO:47 and fragments thereof.

The present invention provides a composition comprising the protein SEQ ID NO:2 encoded by the nucleotide sequence SEQ ID NO:1. The present invention provides a composition comprising the protein SEQ ID NO:4 encoded by the nucleotide sequence SEQ ID NO:3. The present invention provides a composition comprising the protein SEQ ID NO:21 encoded by the nucleotide sequence SEQ ID NO:22. The present invention provides a composition comprising the protein

SEQ ID NO:42 encoded by the nucleotide sequence SEQ ID NO:41. The present invention provides a composition comprising the protein SEQ ID NO:46 encoded by the nucleotide sequence SEQ ID NO:45. The present invention provides a composition comprising the protein SEQ ID NO:48 encoded by the nucleotide sequence SEQ ID NO:47.

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The present invention provides a composition comprising the protein SEQ ID NO:2 or fragments thereof, encoded by the nucleotide sequence SEQ ID NO:1 or fragments thereof. The present invention also provides a composition comprising the protein SEQ ID NO:4 or fragments thereof, encoded by the nucleotide sequence SEQ ID NO:3 or fragments thereof. The present invention also provides a composition comprising the protein SEQ ID NO:21 or fragments thereof, encoded by the nucleotide sequence SEQ ID NO:22 or fragments thereof. The present invention also provides a composition comprising the protein SEQ ID NO:42 or fragments thereof, encoded by the nucleotide sequence SEQ ID NO:41 or fragments thereof. The present invention also provides a composition comprising the protein SEQ ID NO:46 or fragments thereof, encoded by the nucleotide sequence SEQ ID NO:45 or fragments thereof. The present invention also provides a composition comprising the protein SEQ ID NO:48 or fragments thereof, encoded by the nucleotide sequence SEQ ID NO:47 or fragments thereof.

The present invention also provides vectors containing the nucleotide sequences SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:22, SEQ ID NO:41, SEQ ID NO:45, SEQ ID NO:47 or fragments thereof. The present invention also provides cells transfected with these vectors. In addition, the present invention provides cells stably transfected with the nucleotide sequence SEQ ID NO:1 or fragments thereof. The present invention also provides cells stably transfected with the nucleotide sequence SEQ ID NO:3 or fragments thereof. The present invention also provides cells stably transfected with the nucleotide sequence SEQ ID NO:3 or fragments thereof. The present invention also provides cells stably transfected with the nucleotide sequence SEQ ID NO:22 or fragments thereof. The

present invention also provides cells stably transfected with the nucleotide sequence SEQ ID NO:41 or fragments thereof. The present invention also provides cells stably transfected with the nucleotide sequence SEQ ID NO:45 or fragments thereof. The present invention also provides cells stably transfected with the nucleotide sequence SEQ ID NO:47 or fragments thereof.

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The present invention provides cells stably transfected with the nucleotide sequence SEQ ID NO:1 or fragments thereof, which produce the protein SEQ ID NO:2 or fragments thereof. In addition, the present invention provides cells stably transfected with the nucleotide sequence SEQ ID NO:3 or fragments thereof which produce the protein SEQ ID NO:4 or fragments thereof. In addition, the present invention provides cells stably transfected with the nucleotide sequence SEQ ID NO:22 or fragments thereof which produce the protein SEQ ID NO:21 or fragments thereof. The present invention also provides cells stably transfected with the nucleotide sequence SEQ ID NO:41 or fragments thereof which produce the protein SEQ ID NO:42 or fragments thereof. The present invention also provides cells stably transfected with the nucleotide sequence SEQ ID NO:45 or fragments thereof which produce the protein SEQ ID NO:46 or fragments thereof. The present invention also provides cells stably transfected with the nucleotide sequence SEQ ID NO:47 or fragments thereof which produce the protein SEQ ID NO:48 or fragments thereof.

The present invention provides a method for stimulating growth by administering cells stably transfected with the nucleotide sequence SEQ ID NO:1 which produce the protein SEQ ID NO:2 or fragments thereof. The present invention also provides a method for stimulating growth by administering cells stably transfected with the nucleotide sequence SEQ ID NO:3 or fragments thereof, which produce the protein SEQ ID NO:4 or fragments thereof. The present invention also provides a method for stimulating growth by administering cells stably transfected with the nucleotide sequence SEQ ID NO:22 or fragments thereof, which produce

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the protein SEQ ID NO:21 or fragments thereof. The present invention also provides a method for stimulating growth by administering cells stably transfected with the nucleotide sequence SEQ ID NO:41 or fragments thereof, which produce the protein SEQ ID NO:42 or fragments thereof. The present invention also provides a method for stimulating growth by administering cells stably transfected with the nucleotide sequence SEQ ID NO:45 or fragments thereof, which produce the protein SEQ ID NO:46 or fragments thereof. The present invention also provides a method for stimulating growth by administering cells stably transfected with the nucleotide sequence SEQ ID NO:47 or fragments thereof, which produce the protein SEQ ID NO:48 or fragments thereof.

Specifically, the present invention provides a method for stimulating tumor formation by administering cells stably transfected with the nucleotide sequence SEQ ID NO:1 or fragments thereof, which produce the protein SEQ ID NO:2 or fragments thereof. The present invention also provides a method for stimulating tumor formation by administering cells stably transfected with the nucleotide sequence SEQ ID NO:3 or fragments thereof, which produce the protein SEQ ID NO:4 or fragments thereof. The present invention also provides a method for stimulating tumor formation by administering cells stably transfected with the nucleotide sequence SEQ ID NO:22 or fragments thereof, which produce the protein SEQ ID NO:21 or fragments thereof. The present invention also provides a method for stimulating tumor formation by administering cells stably transfected with the nucleotide sequence SEQ ID NO:41 or fragments thereof, which produce the protein SEQ ID NO:42 or fragments thereof. The present invention also provides a method for stimulating tumor formation by administering cells stably transfected with the nucleotide sequence SEQ ID NO:45 or fragments thereof, which produce the protein SEQ ID NO:46 or fragments thereof. The present invention also provides a method for stimulating tumor formation by administering cells stably transfected with the nucleotide sequence SEQ ID NO:47

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or fragments thereof, which produce the protein SEQ ID NO:48 or fragments thereof.

The present invention may also be used to develop anti-sense nucleotide sequences to SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:22, SEQ ID NO:41, SEQ ID NO:45, SEQ ID NO:47 or fragments thereof. These anti-sense molecules may be used to interfere with translation of nucleotide sequences, such as SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:22, SEQ ID NO:41, SEQ ID NO:45, SEQ ID NO:47, or fragments thereof, that encode for proteins such as SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:21, SEQ ID NO:42, SEQ ID NO:46, SEQ ID NO:48 or fragments thereof. Administration of these anti-sense molecules, or vectors encoding for anti sense molecules, to humans and animals, would interfere with production of proteins such as SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:21, SEQ ID NO:42, SEQ ID NO:46, SEQ ID NO:48, or fragments thereof, thereby decreasing production of ROIs and inhibiting cellular proliferation. These methods are useful in producing animal models for use in study of tumor development and vascular growth, and for study of the efficacy of treatments for affecting tumor and vascular growth in vivo.

The present invention also provides a method for high throughput screening of drugs and chemicals which modulate the proliferative activity of the enzymes of the present invention, thereby affecting cell division. Combinatorial chemical libraries may be screened for chemicals which modulate the proliferative activity of these enzymes. Drugs and chemicals may be evaluated based on their ability to modulate the enzymatic activity of the expressed or endogenous proteins, including those represented by SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:21, SEQ ID NO:42, SEQ ID NO:46, SEQ ID NO:48 or fragments thereof. Endogenous proteins may be obtained from many different tissues or cells, such as colon cells. Drugs may also be evaluated based on their ability to bind to the expressed or endogenous proteins represented by SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:21, SEQ ID NO:42, SEQ ID

cytochrome

Expressed proteins or

While not wanting to be

NO:46, SEQ ID NO:48 or fragments thereof. Enzymatic activity may be NADPH- or NADH-dependent superoxide generation catalyzed by the holoprotein. Enzymatic activity may also be NADPH- or NADH-dependent diaphorase activity catalyzed by either the holoprotein or the flavoprotein domain.

the C-terminal half of the enzymes shown in SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:21, SEQ ID NO:42, or fragments thereof, and the C-terminal end of the enzymes shown in SEQ ID NO:46 and SEQ ID NO:48 (approximately the C-terminal

265 amino acids). This fragment of gp91phox has NADPH-

or NADH binding site and the FAD binding site are useful for

activity

By flavoprotein domain, is meant approximately

towards

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nitrobluetetrazolium and other dyes. fragments thereof can be used for robotic screens of existing combinatorial chemical libraries. 15

dependent

bound by the following statement, it is believed that the NADPH

reductase

evaluating the ability of drugs and other compositions to bind to the mox and duox enzymes or to modulate their enzymatic activity. The use of the holoprotein or the C-terminal half or end regions are preferred for developing a high throughput

drug screen. Additionally, the N-terminal one-third of the duox domain (the peroxidase domain) may also be used to evaluate the ability of drugs and other compositions to inhibit the peroxidase activity, and for further development of a high

throughput drug screen.

The present invention also provides antibodies directed to the proteins SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:21, SEQ ID NO:42, SEQ ID NO:46, SEQ ID NO:48 and fragments thereof. The antibodies of the present invention are useful for a variety of purposes including localization, detection and measurement of the proteins SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:21, SEQ ID NO:42, SEQ ID NO:46, SEQ ID NO:48 and fragments thereof. The antibodies may be employed in kits to accomplish these purposes. These antibodies may also be linked to cytotoxic agents for selected killing of cells. The

term antibody is meant to include any class of antibody such as IgG, IgM and other classes. The term antibody also includes a completely intact antibody and also fragments thereof, including but not limited to Fab fragments and Fab + Fc fragments.

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The present invention also provides the nucleotide sequences SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:22, SEQ ID NO:41, SEQ ID NO:45, SEQ ID NO:47 and fragments thereof. These nucleotides are useful for a variety of purposes including localization, detection, and measurement of messenger RNA involved in synthesis of the proteins represented as SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:21, SEQ ID NO:42, SEQ ID NO:46, SEQ ID NO:48 and fragments thereof. nucleotides may also be used in the construction of labeled probes for the localization, detection, and measurement of nucleic acids such as messenger RNA or alternatively for the isolation of larger nucleotide sequences containing nucleotide sequences shown in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:22, SEQ ID NO:41, SEQ ID NO:45, SEQ ID NO:47 or fragments thereof. These nucleotide sequences may be used to isolate homologous strands from other species using techniques known to one of ordinary skill in the art. These nucleotide sequences may also be used to make probes and complementary strands. In particular, the nucleotide sequence shown in SEQ ID NO:47 may be used to isolate the complete coding sequence for duox2. The nucleotides may be employed in kits to accomplish these purposes.

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Most particularly, the present invention involves a method for modulation of growth by modifying the proteins represented as SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:41, SEQ ID NO:42, SEQ ID NO:46, SEQ ID NO:48 or fragments thereof.

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The term "mitogenic regulators" is used herein to mean any molecule that acts to affect cell division.

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The term "animal" is used herein to mean humans and non-human animals of both sexes.

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The terms "a", "an" and "the" as used herein are defined to mean one or more and include the plural unless the context is inappropriate.

"Proteins". "peptides," "polypeptides" "oligopeptides" are chains of amino acids (typically L-amino acids) whose alpha carbons are linked through peptide bonds formed by a condensation reaction between the carboxyl group of the alpha carbon of one amino acid and the amino group of the alpha carbon of another amino acid. The terminal amino acid at one end of the chain (i.e., the amino terminal) has a free amino group, while the terminal amino acid at the other end of the chain (i.e., the carboxy terminal) has a free carboxyl group. As such, the term "amino terminus" (abbreviated N-terminus) refers to the free alpha-amino group on the amino acid at the amino terminal of the protein, or to the alpha-amino group (imino group when participating in a peptide bond) of an amino acid at any other location within the protein. Similarly, the term "carboxy terminus" (abbreviated C-terminus) refers to the free carboxyl group on the amino acid at the carboxy terminus of a protein, or to the carboxyl group of an amino acid at any other location within the protein.

Typically, the amino acids making up a protein are numbered in order, starting at the amino terminal and increasing in the direction toward the carboxy terminal of the protein. Thus, when one amino acid is said to "follow" another, that amino acid is positioned closer to the carboxy terminal of the protein than the preceding amino acid.

The term "residue" is used herein to refer to an amino acid (D or L) or an amino acid mimetic that is incorporated into a protein by an amide bond. As such, the amino acid may be a naturally occurring amino acid or, unless otherwise limited, may encompass known analogs of natural amino acids that function in a manner similar to the naturally occurring amino acids (i.e., amino acid mimetics). Moreover, an amide bond mimetic includes peptide backbone modifications well known to those skilled in the art.

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Furthermore, one of skill will recognize that, as mentioned above, individual substitutions, deletions or additions which alter, add or delete a single amino acid or a small percentage of amino acids (typically less than 5%, more typically less than 1%) in an encoded sequence are conservatively modified variations where the alterations result in the substitution of an amino acid with a chemically similar amino acid. Conservative substitution tables providing functionally similar amino acids are well known in the art. The following six groups each contain amino acids that are conservative substitutions for one another:

- 1) Alanine (A), Serine (S), Threonine (T);
- 2) Aspartic acid (D), Glutamic acid (E);
- 3) Asparagine (N), Glutamine (Q);
- 4) Arginine (R), Lysine (K);
- 5) Isoleucine (I), Leucine (L), Methionine (M), Valine (V); and
- 6) Phenylalanine (F), Tyrosine (Y), Tryptophan (W).

When the peptides are relatively short in length (i.e., less than about 50 amino acids), they are often synthesized using standard chemical peptide synthesis techniques. Solid phase synthesis in which the C-terminal amino acid of the sequence is attached to an insoluble support followed by sequential addition of the remaining amino acids in the sequence is a preferred method for the chemical synthesis of the antigenic epitopes described herein. Techniques for solid phase synthesis are known to those skilled in the art.

Alternatively, the antigenic epitopes described herein are synthesized using recombinant nucleic acid methodology. Generally, this involves creating a nucleic acid sequence that encodes the peptide or protein, placing the nucleic acid in an expression cassette under the control of a particular promoter, expressing the peptide or protein in a host, isolating the expressed peptide or protein and, if required, renaturing the

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peptide or protein. Techniques sufficient to guide one of skill through such procedures are found in the literature.

When several desired protein fragments or peptides are encoded in the nucleotide sequence incorporated into a vector, one of skill in the art will appreciate that the protein fragments or peptides may be separated by a spacer molecule such as, for example, a peptide, consisting of one or more Generally, the spacer will have no specific amino acids. biological activity other than to join the desired protein fragments or peptides together, or to preserve some minimum distance or other spatial relationship between them. However, the constituent amino acids of the spacer may be selected to influence some property of the molecule such as the folding, net charge, or hydrophobicity. Nucleotide sequences encoding for the production of residues which may be useful in purification of the expressed recombinant protein may be built into the vector. Such sequences are known in the art. For example, a nucleotide sequence encoding for a poly histidine sequence may be added to a vector to facilitate purification of the expressed recombinant protein on a nickel column.

Once expressed, recombinant peptides, polypeptides and proteins can be purified according to standard procedures known to one of ordinary skill in the art, including ammonium sulfate precipitation, affinity columns, column chromatography, gel electrophoresis and the like. Substantially pure compositions of about 50 to 99% homogeneity are preferred, and 80 to 95% or greater homogeneity are most preferred for use as therapeutic agents.

One of skill in the art will recognize that after chemical synthesis, biological expression or purification, the desired proteins, fragments thereof and peptides may possess a conformation substantially different than the native conformations of the proteins, fragments thereof and peptides. In this case, it is often necessary to denature and reduce protein and then to cause the protein to re-fold into the preferred conformation. Methods of reducing and denaturing proteins

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and inducing re-folding are well known to those of skill in the art.

The genetic constructs of the present invention include coding sequences for different proteins, fragments thereof, and peptides. The genetic constructs also include epitopes or domains chosen to permit purification or detection of the expressed protein. Such epitopes or domains include DNA sequences encoding the glutathione binding domain from glutathione S-transferase. hexa-histidine, thioredoxin, hemagglutinin antigen, maltose binding protein, and others commonly known to one of skill in the art. The preferred genetic construct includes the nucleotide sequences of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:22, SEQ ID NO:41, SEQ ID NO:45, SEQ ID NO:47 or fragments thereof. It is to be understood that additional or alternative nucleotide sequences may be included in the genetic constructs in order to encode for the following: a) multiple copies of the desired proteins, fragments thereof, or peptides; b) various combinations of the desired proteins, fragments thereof, or peptides; and c) conservative modifications of the desired proteins, fragments thereof, or peptides, and combinations thereof. Preferred proteins include the human mox1 protein and human mox2 protein shown as SEQ ID NO:2 and SEQ ID NO:4, respectively, and fragments thereof. Some preferred fragments of the human mox1 protein (SEQ ID NO:2) include but are not limited to the proteins shown as SEQ ID NO:23, SEQ ID NO:24, and SEQ ID The protein mox1 is also called p65mox in this application. Another preferred protein of the present invention is rat mox1 protein shown as SEQ ID NO:21 and fragments thereof. Another preferred protein of the present invention is rat mox1B protein shown as SEQ ID NO:42 and fragments thereof. Yet another preferred protein of the present invention is duox1 protein shown as SEQ ID NO:46 and fragments thereof. Still another preferred protein of the present invention is duox2 protein. A partial amino acid sequence of the duox2 protein is shown as SEQ ID NO:48.

The nucleotide sequences of the present invention may also be employed to hybridize to nucleic acids such as DNA or RNA nucleotide sequences under high stringency conditions which permit detection, for example, of alternately spliced messages.

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The genetic construct is expressed in an expression system such as in NIH 3T3 cells using recombinant sequences in a pcDNA-3 vector (Invitrogen, Carlsbad, CA) to produce a recombinant protein. Preferred expression systems include but are not limited to Cos-7 cells, insect cells using recombinant baculovirus, and yeast. It is to be understood that other expression systems known to one of skill in the art may be used for expression of the genetic constructs of the present invention. The preferred proteins of the present invention are the proteins referred to herein as human mox1 and human mox2 or fragments thereof which have the amino acid sequences set forth in SEQ ID NO:3 and SEQ ID NO:4, respectively, or an amino acid sequence having amino acid substitutions as defined in the definitions that do not significantly alter the function of the recombinant protein in an adverse manner. Another preferred protein of the present invention is referred to herein as rat mox1 and has the amino acid sequence set forth in SEQ ID NO:21. Yet another preferred protein of the present invention is referred to herein as rat mox1B and has the amino acid sequence set forth in SEQ ID NO:42. Two other preferred proteins of the present invention are referred to herein as human duox1 and human duox2, or fragments thereof, which have the amino acid sequences set forth in SEQ ID NO:46 and SEQ ID NO:48, respectively, or an amino acid sequence having amino acid substitutions as defined in the definitions that do not significantly alter the function of the recombinant protein in an adverse manner.

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Terminology

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It should be understood that some of the terminology used to describe the novel mox and duox proteins

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contained herein is different from the terminology in U.S. Provisional Application Serial No. 60/107,911 and U.S. Provisional Application Serial No. 60/149,332 upon which this application claims priority in part. As described herein, the term "human mox1" refers to a protein comprising an amino acid sequence as set forth in SEQ ID NO:2, or -a fragment thereof, and encoded by the nucleotide sequence as set forth in SEQ ID NO:1, or a fragment thereof. As described herein, the term "human mox2" refers to a protein comprising an amino acid sequence as set forth in SEQ ID NO:4, or a fragment thereof, and encoded by the nucleotide sequence as set forth in SEQ ID NO:3, or a fragment thereof. As described herein, the term "human duox1" refers to a protein comprising an amino acid sequence as set forth in SEQ ID NO:46, or a fragment thereof, and encoded by the nucleotide sequence as set forth in SEQ ID NO:45, or a fragment thereof. As described herein, the term "human duox2" refers to a protein comprising an amino acid sequence as set forth in SEQ ID NO:48, or a fragment thereof, and encoded by the nucleotide sequence as set forth in SEQ ID NO:47, or a fragment thereof.

Construction of the Recombinant Gene

The desired gene is ligated into a transfer vector, such as pcDNA3, and the recombinants are used to transform host cells such as Cos-7 cells. It is to be understood that different transfer vectors, host cells, and transfection methods may be employed as commonly known to one of ordinary skill in the art. Six desired genes for use in transfection are shown in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:22, SEQ ID NO:41 SEQ ID NO:45 and SEQ ID NO:47. For example, lipofectamine-mediated transfection and *in vivo* homologous recombination was used to introduce the mox1 gene into NIH 3T3 cells.

The synthetic gene is cloned and the recombinant construct containing mox or duox gene is produced and grown in confluent monolayer cultures of a Cos-7 cell line. The

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expressed recombinant protein is then purified, preferably using affinity chromatography techniques, and its purity and specificity determined by known methods.

A variety of expression systems may be employed for expression of the recombinant protein. Such expression methods include, but are not limited to the following: bacterial expression systems, including those utilizing *E. coli* and *Bacillus subtilis*; virus systems; yeast expression systems; cultured insect and mammalian cells; and other expression systems known to one of ordinary skill in the art.

Transfection of Cells

It is to be understood that the vectors of the present invention may be transfected into any desired cell or cell line. Both in vivo and in vitro transfection of cells are contemplated as part of the present invention. Preferred cells for transfection include but are not limited to the following: fibroblasts (possibly to enhance wound healing and skin formation), granulocytes (possible benefit to increase function in a compromised immune system as seen in AIDS, and aplastic anemia), muscle cells, neuroblasts, stem cells, bone marrow cells, osteoblasts, B lymphocytes, and T lymphocytes.

Cells may be transfected with a variety of methods known to one of ordinary skill in the art and include but are not limited to the following: electroporation, gene gun, calcium phosphate, lipofectamine, and fugene, as well as adenoviral transfection systems.

Host cells transfected with the nucleic acids represented in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:22, SEQ ID NO:41 SEQ ID NO:45 and SEQ ID NO:47, or fragments thereof, are used to express the proteins SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:21, SEQ ID NO:42, SEQ ID NO:46 and SEQ ID NO:48, respectively, or fragments thereof.

These expressed proteins are used to raise antibodies. These antibodies may be used for a variety of applications including but not limited to immunotherapy against

cancers expressing one of the mox or duox proteins, and for detection, localization and measurement of the proteins shown in SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:21, SEQ ID NO:42, SEQ ID NO:48 or fragments thereof.

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Purification and Characterization of the Expressed Protein

The proteins of the present invention can be expressed as a fusion protein with a poly histidine component, such as a hexa histidine, and purified by binding to a metal affinity column using nickel or cobalt affinity matrices. protein can also be expressed as a fusion protein with glutathione S-transferase and purified by affinity chromatography using a glutathione agarose matrix. The protein can also be purified by immunoaffinity chromatography by expressing it as a fusion protein, for example with hemagglutinin antigen. The expressed or naturally occurring protein can also be purified by conventional chromatographic and purification methods which include anion and cation exchange chromatography, gel exclusion chromatography, hydroxylapatite chromatography, dye binding chromatography, ammonium sulfate precipitation, precipitation in organic solvents or other techniques commonly known to one of skill in the art.

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Methods of Assessing Activity of Expressed Proteins

Different methods are available for assessing the activity of the expressed proteins of the present invention, including but not limited to the proteins represented as SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:21, SEQ ID NO:42, SEQ ID NO:46 or SEQ ID NO:48 substituted analogs thereof, and fragments thereof.

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1. Assays of the holoprotein and fragments thereof for superoxide generation:

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A. General considerations.

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These assays are useful in assessing efficacy of drugs designed to modulate the activity of the enzymes of the present invention. The holoprotein may be expressed in COS-7 cells, NIH 3T3 cells, insect cells (using baculoviral technology) or other cells using methods known to one of skill in the art. Membrane fractions or purified protein are used for the assay. The assay may require or be augmented by other cellular proteins such as p47phox, p67phox, and Rac1, as well as potentially other unidentified factors (e.g., kinases or other regulatory proteins).

B. Cytochrome c reduction.

NADPH or NADH is used as the reducing substrate, in a concentration of about 100 µM. Reduction of cytochrome c is monitored spectrophotometrically by the increase in absorbance at 550 nm, assuming an extinction coefficient of 21 mM⁻¹cm⁻¹. The assay is performed in the absence and presence of about 10 µg superoxide dismutase. The superoxide-dependent reduction is defined as cytochrome c reduction in the absence of superoxide dismutase minus that in the presence of superoxide dismutase (Uhlinger et al. (1991) J. Biol. Chem. 266, 20990-20997). Acetylated cytochrome c may also be used, since the reduction of acetylated cytochrome c is thought to be exclusively via superoxide.

C. Nitroblue tetrazolium reduction.

For nitroblue tetrazolium (NBT) reduction, the same general protocol is used, except that NBT is used in place of cytochrome c. In general, about 1 mL of filtered 0.25 % nitrotetrazolium blue (Sigma, St. Louis, MO) is added in Hanks buffer without or with about 600 Units of superoxide dismutase (Sigma) and samples are incubated at approximately 37°C. The oxidized NBT is clear, while the reduced NBT is blue and insoluble. The insoluble product is collected by centrifugation, and the pellet is re-suspended in about 1 mL of pyridine (Sigma) and heated for about 10 minutes at 100°C to solubilize

the reduced NBT. The concentration of reduced NBT is determined by measuring the absorbance at 510 nm, using an extinction coefficient of 11,000 M⁻¹cm⁻¹. Untreated wells are used to determine cell number.

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D. Luminescence.

Superoxide generation may also be monitored with a chemiluminescence detection system utilizing lucigenin (bis-N-methylacridinium nitrate, Sigma, St. Louis, MO). The sample is mixed with about 100 µM NADPH (Sigma, St. Louis, MO) and 10 µM lucigenin (Sigma, St. Louis, MO) in a volume of about 150 µL Hanks solution. Luminescence is monitored in a 96-well plate using a LumiCounter (Packard, Downers Grove, IL) for 0.5 second per reading at approximately 1 minute intervals for a total of about 5 minutes; the highest stable value in each data set is used for comparisons. As above, superoxide dismutase is added to some samples to prove that the luminescence arises from superoxide. A buffer blank is subtracted from each reading (Ushio-Fukai et al. (1996) J. Biol. Chem. 271, 23317-23321).

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E. Assays in intact cells.

Assays for superoxide generation may be performed using intact cells, for example, the mox-transfected NIH 3T3 cells. In principle, any of the above assays can be used to evaluate superoxide generation using intact cells, for example, the mox-transfected NIH 3T3 cells. NBT reduction is a preferred assay method.

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2. Assays of truncated proteins comprised of approximately the C-terminal 265 amino acid residues

While not wanting to be bound by the following statement, the truncated protein comprised of approximately the C-terminal 265 amino acid residues is not expected to generate superoxide, and therefore, superoxide dismutase is not added in assays of the truncated protein. Basically, a similar assay is

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established and the superoxide-independent reduction of NBT, cytochrome c, dichlorophenolindophenol, ferricyanide, or another redox-active dye is examined.

Nucleotides and Nucleic Acid Probes

The nucleotide sequences SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:22, SEQ ID NO:41 SEQ ID NO:45 and SEQ ID NO:47, as well as fragments thereof and PCR primers therefor, may be used, respectively, for localization, detection and measurement of nucleic acids related to SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:22, SEQ ID NO:41 SEQ ID NO:45 and SEQ ID NO:47, as well as fragments thereof. The nucleotide sequences SEQ ID NO:1 and SEQ ID NO:3 are also called the human mox1 gene and the human mox2 gene in this application. SEQ ID NO:41 is also known as the rat mox1B gene in this application. SEQ ID NO:45 is also known as the human duox1 gene in this application. SEQ ID NO:47 is also known as the human duox1 gene in this application. SEQ ID NO:47 is also known as the human duox2 gene in this application.

The nucleotide sequences SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:22, SEQ ID NO:41 SEQ ID NO:45 and SEQ ID NO:47, as well as fragments thereof, may be used to create probes to isolate larger nucleotide sequences containing the nucleotide sequences SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:22, SEQ ID NO:41 SEQ ID NO:45 and SEQ ID NO:47, respectively. The nucleotide sequences SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:22, SEQ ID NO:41 SEQ ID NO:45 and SEQ ID NO:47, as well as fragments thereof, may also be used to create probes to identify and isolate mox and duox proteins in other species.

The nucleic acids described herein include messenger RNA coding for production of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:45, SEQ ID NO:46, SEQ ID NO:48 and fragments thereof. Such nucleic acids include but are not limited to cDNA probes. These probes may be labeled in a variety of ways known to one of ordinary skill in

the art. Such methods include but are not limited to isotopic and non-isotopic labeling. These probes may be used for *in situ* hybridization for localization of nucleic acids such as mRNA encoding for SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:21, SEQ ID NO:42, SEQ ID NO:46, SEQ ID NO:48 and fragments thereof. Localization may be performed using *in situ* hybridization at both ultrastructural and light microscopic levels of resolution using techniques known to one of ordinary skill in the art.

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These probes may also be employed to detect and quantitate nucleic acids and mRNA levels using techniques known to one of ordinary skill in the art including but not limited to solution hybridization.

15 Antibody Production

The proteins shown in SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:41, SEQ ID NO:42, SEQ ID NO:46, SEQ ID NO:48, or fragments thereof, are combined with a pharmaceutically acceptable carrier or vehicle to produce a pharmaceutical composition and administered to animals for the production of polyclonal antibodies using methods known to one of ordinary skill in the art. The preferred animals for antibody production are rabbits and mice. Other animals may be employed for immunization with these proteins or fragments thereof. Such animals include, but are not limited to the following; sheep, horses, pigs, donkeys, cows, monkeys and rodents such as guinea pigs and rats.

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The terms "pharmaceutically acceptable carrier or pharmaceutically acceptable vehicle" are used herein to mean any liquid including but not limited to water or saline, oil, gel, salve, solvent, diluent, fluid ointment base, liposome, micelle, giant micelle, and the like, which is suitable for use in contact with living animal or human tissue without causing adverse physiological responses, and which does not interact with the other components of the composition in a deleterious manner.

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The pharmaceutical compositions may conveniently be presented in unit dosage form and may be prepared by conventional pharmaceutical techniques. Such techniques include the step of bringing into association the active ingredient and the pharmaceutical carrier(s) or excipient(s). In general, the formulations are prepared by uniformly and intimately bringing into association the active ingredient with liquid carriers.

Formulations suitable for parenteral administration include aqueous and non-aqueous sterile injection solutions which may contain anti-oxidants, buffers, bacteriostats and solutes which render the formulation isotonic with the blood of the intended recipient; and aqueous and non-aqueous sterile suspensions which may include suspending agents and thickening agents. The formulations may be presented in unit-dose or multi-dose containers, for example, sealed ampules and vials, and may be stored in a freeze-dried (lyophilized) condition requiring only the addition of the sterile liquid carrier, for example, water for injections, immediately prior to use. Extemporaneous injection solutions and suspensions may be prepared from sterile powders, granules and tablets commonly used by one of ordinary skill in the art.

Preferred unit dosage formulations are those containing a dose or unit, or an appropriate fraction thereof, of the administered ingredient. It should be understood that in addition to the ingredients, particularly mentioned above, the formulations of the present invention may include other agents commonly used by one of ordinary skill in the art.

The pharmaceutical composition may be administered through different routes, such as oral, including buccal and sublingual, rectal, parenteral, aerosol, nasal, intramuscular, subcutaneous, intradermal, and topical. The pharmaceutical composition of the present invention may be administered in different forms, including but not limited to solutions, emulsions and suspensions, microspheres, particles, microparticles, nanoparticles, and liposomes. It is expected that

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from about 1 to 7 dosages may be required per immunization regimen. Initial injections may range from about 0.1 μ g to 1 mg, with a preferred range of about 1 μ g to 800 μ g, and a more preferred range of from approximately 25 μ g to 500 μ g. Booster injections may range from 0.1 μ g to 1 mg, with a preferred range of approximately 1 μ g to 800 μ g, and a more preferred range of about 10 μ g to 500 μ g.

The volume of administration will vary depending on the route of administration and the size of the recipient. For example, intramuscular injections may range from about 0.1 ml to 1.0 ml.

The pharmaceutical composition may be stored at temperatures of from about 4°C to -100°C. The pharmaceutical composition may also be stored in a lyophilized state at different temperatures including room temperature. The pharmaceutical composition may be sterilized through conventional means known to one of ordinary skill in the art. Such means include, but are not limited to filtration, radiation and heat. The pharmaceutical composition of the present invention may also be combined with bacteriostatic agents, such as thimerosal, to inhibit bacterial growth.

Adjuvants

A variety of adjuvants known to one of ordinary skill in the art may be administered in conjunction with the protein in the pharmaceutical composition. Such adjuvants include, but are not limited to the following: polymers, copolymers such polyoxyethylene-polyoxypropylene as copolymers, including block co-polymers; polymer P1005; Freund's complete adjuvant (for animals); Freund's incomplete adjuvant; sorbitan monooleate; squalene; CRL-8300 adjuvant; alum; QS 21, muramyl dipeptide; trehalose; bacterial extracts, mycobacterial extracts; detoxified endotoxins: membrane lipids; or combinations thereof.

Monoclonal antibodies can be produced using hybridoma technology in accordance with methods well known

to those skilled in the art. The antibodies are useful as research or diagnostic reagents or can be used for passive immunization. The composition may optionally contain an adjuvant.

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The polyclonal and monoclonal antibodies useful as research or diagnostic reagents may be employed for detection and measurement of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:4, SEQ ID NO:45, SEQ ID NO:48 and fragments thereof. Such antibodies may be used to detect these proteins in a biological sample, including but not limited to samples such as cells, cellular extracts, tissues, tissue extracts, biopsies, tumors, and biological fluids. Such detection capability is useful for detection of disease related to these proteins to facilitate diagnosis and prognosis and to suggest possible treatment alternatives.

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Detection may be achieved through the use of immunocytochemistry, ELISA, radioimmunoassay or other assays as commonly known to one of ordinary skill in the art. The mox1, mox2, duox1 and duox2 proteins, or fragments thereof, may be labeled through commonly known approaches, including but not limited to the following: radiolabeling, dyes, magnetic particles, biotin-avidin, fluorescent molecules, chemiluminescent molecules and systems, ferritin, colloidal gold, and other methods known to one of skill in the art of labeling proteins.

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Administration of Antibodies

The antibodies directed to the proteins shown as SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:21, SEQ ID NO:42, SEQ ID NO:46 or SEQ ID NO:48, or directed to fragments thereof, may also be administered directly to humans and animals in a passive immunization paradigm. Antibodies directed to extracellular portions of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:41, SEQ ID NO:42, SEQ ID NO:46 or SEQ ID NO:48 bind to these extracellular epitopes. Attachment of labels to these antibodies facilitates localization and visualization of sites of binding. Attachment of molecules such as ricin or

other cytotoxins to these antibodies helps to selectively damage or kill cells expressing SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:21, SEQ ID NO:42, SEQ ID NO:46, SEQ ID NO:48 or fragments thereof.

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Kits

The present invention includes kits useful with the antibodies, nucleic acids, nucleic acid probes, labeled antibodies, labeled proteins or fragments thereof for detection, localization and measurement of SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:21, SEQ ID NO:22, SEQ ID NO:41, SEQ ID NO:42, SEQ ID NO:45, SEQ ID NO:46, SEQ ID NO:47, SEQ ID NO:48 or combinations and fragments thereof.

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Kits may be used for immunocytochemistry, in situ hybridization, solution hybridization, radioimmunoassay, ELISA, Western blots, quantitative PCR, and other assays for the detection, localization and measurement of these nucleic acids, proteins or fragments thereof using techniques known to one of skill in the art.

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The nucleotide sequences shown in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:22, SEQ ID NO:41 SEQ ID NO:45, SEQ ID NO:47, or fragments thereof, may also be used under high stringency conditions to detect alternately spliced messages related to SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:22, SEQ ID NO:41 SEQ ID NO:45, SEQ ID NO:47 or fragments thereof, respectively.

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As discussed in one of the Examples, rat mox1 protein (SEQ ID NO: 21) is similar to mouse gp91 protein (SEQ ID NO: 38), whereas rat mox1B protein (SEQ ID NO:42) is similar to human gp91 protein (SEQ ID NO:12). This observation suggests that other isoforms of mouse and human gp91 may exist. In addition, another subtype of human mox1, similar to rat mox1B (SEQ ID NO:42), also exists. The presence of two isoforms of rat mox1 protein in vascular smooth muscle may have important physiological consequences

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and biomedical applications. For example, the two isoforms may have different biological activities, different tissue distributions and may be regulated differently in physiological and/or pathological conditions. The fact that mox1B (SEQ ID NO:42) was isolated from cells exposed to angiotensin II, known to promote oxidative stress and vascular growth, suggests that it may be upregulated by this hormone and may be overexpressed in disease. Therefore, the diagnostic kits of the present invention can measure the relative expression of the two mox isoforms. The diagnostic kits may also measure or detect the relative expression of the mox proteins described herein (i.e. human mox1 and/or human mox2) and duox proteins described herein (i.e. human duox1 and/or human duox2).

Fragments of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:22, SEQ ID NO:41 SEQ ID NO:45 and SEQ ID NO:47 containing the relevant hybridizing sequence can be synthesized onto the surface of a chip array. RNA samples, e.g., from tumors, are then fluorescently tagged and hybridized onto the chip for detection. This approach may be used diagnostically to characterize tumor types and to tailor treatments and/or provide prognostic information. Such prognostic information may have predictive value concerning disease progression and life span, and may also affect choice of therapy.

The present invention is further illustrated by the following examples, which are not to be construed in any way as imposing limitations upon the scope thereof. On the contrary, it is to be clearly understood that resort may be had to various other embodiments, modifications, and equivalents thereof, which, after reading the description herein, may suggest themselves to those skilled in the art without departing from the spirit of the present invention.

EXAMPLE 1

Sequence Analysis and Cloning of the Human mox1 cDNA (SEQ ID NO:1) Encoding for Production of the Human mox1 Protein p65mox (SEQ ID NO:2)

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Using gp91phox as a query sequence, a 334 base sequenced portion of expressed sequence tag (EST) 176696 (GenBank Accession number AA305700) showed 68.8% sequence identity at the predicted amino acid level with human (h) gp91phox. The bacterial strain number 129134 containing the EST sequence in the pBluescript SK- vector, was purchased from American Tissue Type Culture Collection (ATCC, Rockville, MD). The EST sequence was originally cloned from a Caco-2 human colon carcinoma cell line. The EST176696 DNA was further sequenced using the T7 and T3 vector promoters and primers designed to match the known 3'

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Internal primers used for sequencing were as sequence. follows: 5'-AAC AAG CGT GGC TTC AGC ATG-3' SEQ ID NO:5 (251S, numbering is based on the nucleotides from the 5' end of EST176696, and S indicates the sense direction), 5'-AGC

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AAT ATT GTT GGT CAT-3' SEQ ID NO:6 (336S), 5'-GAC TTG ACA GAA AAT CTA TAA GGG-3' SEQ ID NO:7 (393S), 5'-TTG TAC CAG ATG GAT TTC AA-3' SEQ ID NO:8 (673A, A indicates the antisense direction), 5'-CAG GTC TGA AAC AGA AAA CCT-3' SEQ ID NO:9 (829S), 5'-ATG

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AAT TCT CAT TAA TTA TTC AAT AAA-3' SEQ ID NO:10 The coding sequence in EST176696 showed (1455A). homology to a 250 amino acid stretch corresponding to the Nterminal 44% of human gp91phox, and contained a stop codon

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corresponding to the location in human gp91phox. 5' Rapid amplification of cDNA ends (RACE) was carried out using a human colon cDNA library and Marathon cDNA Amplification Kit (ClonTech) using 5'-ATC TCA AAA GAC TCT GCA CA-3' SEQ ID NO:11 (41A) as an internal gene-specific primer (Frohman et al. (1988) Proc. Natl. Acad. Sci. USA 85, 8998-

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9002). 5' RACE resulted in a 1.1 kb fragment representing the complete 5' sequence, based on homology with gp91phox.

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Reamplification was performed with primers spanning the putative start and stop codons, using the 1.1 kb 5' RACE product and pSK-EST176696 for primer design. The amplified 1.7kb fragment was TA cloned into the PCR2.1 vector (Invitrogen, Carlsbad, CA). This recombinant vector is referred to as PCR-mox.

Figure 1(a-d) presents a comparison of the present amino acid sequences of human, bovine and murine gp91 phox with the human and rat mox1 proteins of the present invention and the human duox2 protein of the present invention. Also shown are the amino acid sequences for related plant enzyme proteins.

The encoded hp65mox ("mox" referring to mitogenic oxidase and "65" referring to its predicted molecular weight) is listed as SEQ ID NO:2. h-gp91phox (SEQ ID NO:12) and SEQ ID NO:2 differ in length by 3 residues and are 70% identical in their amino acid sequence. h-gp91phox and SEQ ID NO:2 show a greater percentage identity in the C-terminal half of the molecule which contains the putative NADPH and FAD binding sites, and there are several relatively long stretches of complete identity within this region.

A dendrogram (Figure 2) comparing the amino acid sequences of mouse and human gp91phox with that of mox1 SEQ ID NO:2 shows that the latter probably represents a distinct isoform of gp91phox. Two plant homologs of cytochrome b₅₅₈ large subunit are also indicated and represent more distant relatives of the human sequences. Human (and rat mox1 described more fully below) lack asparagine-linked glycosylation sites, which are seen in the highly glycosylated human and mouse gp91phox. Additionally, the hydropathy profiles of human gp91phox and mox1 are nearly identical and include five very hydrophobic stretches in the amino-terminal half of the molecules which are predicted to be membrane-spanning regions.

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EXAMPLE 2

Expression of Mox1

Human multiple tissue northern (MTN) Blot I and Human MTN Blot IV (ClonTech) membranes were hybridized with the putative coding region of the PCR-mox vector at 68° C for several hours. The mox coding region was labeled by random priming with $[\alpha^{-32}P]dCTP$ (10 μ Ci) using the Prime-It II kit (Stratagene). For analysis of mox1 expression in cell lines, total RNA was prepared from 10^{6} cells using the High Pure RNA Isolation Kit (Boehringer Mannheim) or RNeasy kit (Quiagen). Total RNA (10-20 μ g) was separated on a 1% agarose formaldehyde mini-gel and transferred to a Nytran filter (Biorad) and immobilized by ultraviolet cross-linking.

Northern blotting revealed that the major location of mRNA coding for the mox1 protein was colon. The message was also detected in prostate and uterus. The human colon-carcinoma cell line, Caco-2, also expressed large quantities of mox1 message. Northern blotting of mRNA from rat aortic smooth muscle cells also showed strong hybridization, which increased roughly two-fold within 12 hours after treatment with platelet-derived growth factor. This increase in the expression of rat mox1 is consistent with the idea that mox1 contributes to the growth-stimulatory effects of PDGF.

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EXAMPLE 3

Transfection of NIH3T3 Cells with SEQ ID NO:1

The nucleotide sequence (SEQ ID NO:1) encoding for production of the mox1 protein (SEQ ID NO:2) was subcloned into the *Not1* site of the pEF-PAC vector (obtained from Mary Dinauer, Indiana University Medical School, Indianapolis, IN) which has a puromycin resistance gene. Transfection was carried out as described in Sambrook et al., Molecular Cloning, A Laboratory Manual, Volumes 1-3, 2nd edition, Cold Spring Harbor Laboratory Press, N.Y., 1989. The SEQ ID NO:1 in pEF-PAC and the empty vector were separately transfected into NIH 3T3 cells using Fugene 6

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(Boeringer Mannheim). About 2 x 10^6 cells maintained in DMEM containing 10% calf serum were transfected with 10 μ g of DNA. After 2 days, cells were split and selected in the same medium containing 1mg/ml puromycin. Colonies that survived in selection media for 10 to 14 days were subcultured continuously in the presence of puromycin.

Transfected cells exhibited a "transformed"-like morphology, similar to that seen with (V12)Ras-transfected cells, characterized by long spindle-like cells. The parent NIH 3T3 cells or cells transfected with the empty vector showed a normal fibroblast-like morphology.

EXAMPLE 4

Expression of Mox1 (SEQ ID NO:1) in Transfected NIH3T3 Cells

To verify the expression of mox1 mRNA after transfection, RT-PCR and Northern blotting were performed. Total RNAs were prepared from 106 cells using the High Pure RNA Isolation Kit (Boeringer Mannheim) or RNeasy kit (Qiagen). cDNAs for each colony were prepared from 1-2 µg of total RNA using Advantage RT-PCR Kit (ClonTech). PCR amplification was performed using primers, 5'-TTG GCT AAA TCC CAT CCA-3' SEQ ID NO:13 (NN459S, numbering containing NN indicates numbering from the start codon of mox1) and 5'-TGC ATG ACC AAC AAT ATT GCT G-3' SEQ ID NO:14 (NN1435A). For Northern blotting, 10-20 µg of total RNA was separated on a 1% agarose formaldehyde gel and transferred to a nylon filter. After ultraviolet (UV) crosslinking, filters were used for Northern blotting assay as described in Example 2.

Colonies expressing large amounts of mox1 mRNA were chosen for further analysis. The expression of mRNA for glyceraldehyde 3 phosphate dehydrogenase in the various cell lines was normal.

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Colony Formation on Soft Agar

10⁵ to 10³ cells stably transfected with human mox1 gene SEQ ID NO:1 and with empty vector were prepared in 0.3% warm (40°C) agar solution containing DMEM and 10% calf serum. Cells were distributed onto a hardened 0.6% agar plate prepared with DMEM and 10% calf serum. After three weeks in culture (37°C, 5% CO₂) colony formation was observed by microscopy.

Cells which were stably transfected with the empty vector and cultured in soft agar for 3 weeks as above did not display anchorage independent growth. In contrast, NIH 3T3 cells which had been stably transfected with the mox1 (SEQ ID NO:1) and cultured for 3 weeks in soft agar demonstrated anchorage independent growth of colonies.

anchorage independent growth of colonie

EXAMPLE 6

NADPH-Dependent Superoxide Generation Assay

In one embodiment of the present invention, NIH 3T3 cells stably transfected with the human mox1 gene (SEQ ID NO:1) were analyzed for superoxide generation using the lucigenin (Bis-N-methylacridinium luminescence assay (Sigma, St. Louis, MO, Li et al. (1998) *J. Biol. Chem.* 273, 2015-2023). Cells were washed with cold HANKS' solution and homogenized on ice in HANKS' buffer containing 15% sucrose using a Dounce homogenizer. Cell lysates were frozen immediately in a dry ice/ethanol bath. For the assay, 30 µg of cell lysate was mixed with 200 µM NADPH and 500 µM lucigenin. Luminescence was monitored using a LumiCounter (Packard) at three successive one minute intervals and the highest value was used for comparison. Protein concentration was determined by the Bradford method.

Superoxide generation was monitored in lysates from some of the stably transfected cell lines and was compared with superoxide generation by the untransfected NIH 3T3 cell lysates. The results are shown in Table 4. Cell lines 26, 27, and 28 gave the highest degree of morphological changes by

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microscopic examination corresponding to the highest degree of superoxide generation. The luminescent signal was inhibited by superoxide dismutase and the general flavoprotein inhibitor diphenylene iodonium, but was unaffected by added recombinant human p47phox, p67phox and Rac1(GTP-γS), which are essential cytosolic factors for the phagocyte respiratory-burst oxidase.

Table 4

	Table 4		
10	Cell Line Name	Superoxide Generation	
		(RLU)	
	Control (untransfected)	6045	
	mox1-26	17027	
	mox1-27	14670	
15	mox1-28	18411	
	mox1-65	5431	
	mox1-615	11331	
	mox1-+3	8645	
	mox1-+10	5425	
20	mox1-pcc16	8050	

In an alternate and preferred embodiment of the present invention, cells that had been stably transfected with mox1 (YA28) or with empty vector (NEF2) were grown in 10 cm tissue culture plates in medium containing DMEM, 10% calf serum, 100 units/ml penicillin, 100 μ g/ml streptomycin, and 1 μ g/ml puromycin to approximately 80% confluency. Cells (five tissue culture plates of each cell type) were washed briefly with 5 ml phosphate buffered saline (PBS) then dissociated from the plates with PBS containing 5 mM EDTA. Cells were pelleted by centrifuging briefly at 1000 x g.

To permeabilize the cells, freeze thaw lysis was carried out and this was followed by passage of the cell material through a small bore needle. The supernatant was removed and the cells were frozen on dry ice for 15 minutes. After cells were thawed, 200 µl lysis buffer (HANKS' Buffered Salt

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Solution - HBBS) containing a mixture of protease inhibitors from Sigma (Catalog # P2714) was added. Cells on ice were passed through an 18 guage needle 10 times and 200 µl of HBSS buffer containing 34% sucrose was added to yield a final concentration of 17% sucrose. Sucrose appeared to enhance stability upon storage. The combination of freeze-thawing and passage through a needle results in lysis of essentially all of the cells, and this material is referred to as the "cell lysate."

The cell lysates were assayed for protein concentration using the BioRad protein assay system. Cell lysates were assayed for NADPH-dependent chemiluminescence combining HBSS buffer, arachidonic acid, and $0.01-1~\mu g$ protein in assay plates (96 well plastic plates). The reaction was initiated by adding 1.5 mM NADPH and 75 µM lucigenin to the assay mix to give a final concentration of 200 µM NADPH and 10 µM lucigenin, and the chemiluminescence was monitored immediately. The final assay volume as 150 µl. The optimal arachidonic acid concentration was between 50-100 µM. Packard Lumicount luminometer was used to measure chemiluminescence of the reaction between lucigenin and superoxide at 37°C. The plate was monitored continuously for 60 minutes and the maximal relative luminescence unit (RLU) value for each sample was used for the graph.

Figure 3 shows the RLU at various concentrations of cell lysates from mox1-transfected (YA28) and vector control (NEF2) cells. The presence of NaCl or KCl within a concentration range of 50-150 µM is important for optimal activity. MgCl₂(1-5 mM) further enhanced activity by about 2-fold. This cell-free assay for mox1 NADPH-oxidase activity is useful for screening modulators (inhibitors or stimulators) of the mox1 enzyme. The assay may also be used to detect mox and duox NADPH-oxidase activity in general and to screen for modulators (inhibitors or stimulators) of the mox and duox family of enzymes.

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EXAMPLE 7

Nitro Blue Tetrazolium Reduction by Superoxide Generated by NIH 3T3 cells Transfected with the Mox1 cDNA (SEQ ID NO:1)

Superoxide generation by intact cells monitored by using superoxide dismutase-sensitive reduction of nitroblue tetrazolium. NEF2 (vector alone control), YA26 (mox1 (SEQ ID NO:1)-transfected) and YA28 (mox1 (SEQ ID NO:1)-transfected) cells were plated in six well plates at 500,000 cells per well. About 24 hours later, medium was removed from cells and the cells were washed once with 1 mL Hanks solution (Sigma, St. Louis, MO). About 1 mL of filtered 0.25% Nitro blue tetrazolium (NBT, Sigma) was added in Hanks without or with 600 units of superoxide dismutase (Sigma) and cells were incubated at 37°C in the presence of 5% CO₂. After 8 minutes the cells were scraped and pelleted at more than 10,000g. The pellet was re-suspended in 1 mL of pyridine (Sigma) and heated for 10 minutes at 100°C to solubilize the reduced NBT. The concentration of reduced NBT was determined by measuring the absorbance at 510 nm, using an extinction coefficient of 11,000 M-1cm-1. Some wells were untreated and used to determine cell number.

The data are presented in Table 5 and Figure 4 and indicate that the mox1 (SEQ ID NO:1)-transfected cells generated significant quantities of superoxide.

Table 5

-	NBT Reduction (nmols/106 cells)	- SOD	<u>+ SOD</u>
	vector control cells	2.5 ± 0.5	2.1 ± 0.5
30	YA26 (mox1) cells	6.4 ± 0.2	3.4 ± 0.1
	YA28 (mox1) cells	5.2 ± 0.6	3.4 ± 0.3

-SOD, and +SOD mean in the absence or presence of added superoxide dismutase, respectively.

Because superoxide dismutase is not likely to penetrate cells, superoxide must be generated extracellularly. The amount of

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superoxide generated by these cells is about 5-10% of that generated by activated human neutrophils.

EXAMPLE 8

Modification of Intracellular Components in Mox1 Transfected Cells

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To test whether superoxide generated by mox1 can affect intracellular "targets," aconitase activity in control and moxtransfected cell lines was monitored as described in Suh et al. (1999) Nature 401, 79-82. Aconitase contains a four-ironsulphur cluster that is highly susceptible to modification by superoxide, resulting in a loss of activity, and has been used as a reporter of intra-cellular superoxide generation. activity was determined as described in Gardner et al. (1995) J. Biol. Chem. 270, 13399-13405. Acotinase activity was significantly diminished in all three mox-transfected cell lines designated YA26, YA28 and YA212 as compared to the transfected control (Figure 5). Approximately 50% of the aconitase in these cells is mitochondrial, based on differential centrifugation, and the cytosolic and mitochondrial forms were both affected. Control cytosolic and mitochondrial enzymes that do not contain iron-sulfur centres were not affected. Superoxide generated in mox1-transfected cells is therefore capable of reacting with and modifying intracellular components.

EXAMPLE 9

Tumor Generation in Nude Mice Receiving Cells Transfected with the Human mox1 cDNA (SEQ ID NO:1)

About 2 x 10⁶ NIH 3T3 cells (either mox1-transfected with SEQ ID NO:1 or cells transfected using empty vector) were injected subdermally into the lateral aspect of the neck of 4-5 week old nude mice. Three to six mice were injected for each of three mox1-transfected cell lines, and 3 mice were injected with the cells transfected with empty vector (control). After 2 to 3 weeks, mice were sacrificed. The tumors were fixed in 10% formalin and characterized by

histological analysis. Tumors averaged 1.5 x 1 x 1 cm in size and showed histology typical of sarcoma type tumors. addition, tumors appeared to be highly vascularized with superficial capillaries. Eleven of twelve mice injected with mox1 gene-transfected cells developed tumors, while none of the three control animals developed tumors.

In another study, 15 mice were injected with mox1transfected NIH 3T3 cells. Of the 15 mice injected, 14 showed large tumors within 17 days of injection, and tumors showed expression of mox1 mRNA. Histologically, the tumors resembled fibrosarcomas and were similar to ras-induced tumors. Thus, ras and mox1 were similarly potent in their ability to induce tumorigenicity of NIH 3T3 cells in athymic mice.

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EXAMPLE 10

Demonstration of the Role of Mox1 in Non-Cancerous Growth

A role in normal growth was demonstrated in rat aortic vascular smooth-muscle cells by using antisense to rat Transfection with the antisense DNA resulted in a decrease in both superoxide generation and serum-dependent growth. Mox1 is therefore implicated in normal growth in this cell type.

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EXAMPLE 11

Expression of Human Mox1 Protein (SEQ ID NO:2) in a Baculovirus Expression System

SEQ ID NO:2 was also expressed in insect cells using recombinant baculovirus. To establish the p65mox1 expressing virus system, the mox1 gene (SEQ ID NO:1) was initially cloned into the pBacPAK8 vector (Clontech, Palo Alto, CA) and recombinant baculovirus was constructed using standard methods according to manufacturer's protocols. Briefly, PCR amplified mox1 DNA was cloned into the KpnI and EcoRI site of the vector. Primers used for PCR amplification were: 5'-CAA GGT ACC TCT TGA CCA TGG

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GAA ACT-3', SEQ ID NO:15, and 5'-ACG AAT TCA AGT AAA TTA CTG AAG ATA C-3', SEQ ID NO:16. Sf9 insect cells (2 x 106 cells) were infected with 0.5 mg of linearized baculovirus DNA sold under the trademark BACULOGOLD® (PharMingen, San Diego, CA) and 5 mg pBacPAC8-p65mox1 using Transfection Buffers A and B (PharMingen, San Diego, After 5 days, the supernatants containing recombinant viruses were harvested and amplified by infecting fresh sf9 cells for 7 days. Amplification was carried out three times and the presence of the recombinant virus containing mox1 DNA was confirmed by PCR using the same primers. After three times amplification of viruses, plaque purification was carried out to obtain the high titer viruses. Approximately 2 x 108 sf9 cells in agar plates were infected for 5 days with serial dilutions of virus and were dyed with neutral red for easy detection of virus plaques. Selected virus plaques were extracted and the presence of the human mox1 DNA was confirmed again by PCR.

EXAMPLE 12

Cloning of a Rat Homolog of p65mox (SEQ ID NO:2)

cDNA clones of p65mox from a rat aortic smooth muscle cell have been obtained. RT-PCR (reverse transcription polymerase chain reaction) was carried out as follows: first strand cDNA synthesis was performed using total RNA from rat aortic vascular smooth muscle cells, oligo dT primer and superscript II reverse transcriptase, and followed by incubation with RNase H. Degenerate PCR primers were designed to anneal to conserved areas in the coding regions of h-mox1 and gp91phox of human (X04011), mouse (U43384) and porcine (SSU02476) origin. **Primers** were: sense 5'-CCIGTITGTCGIAATCTGCTSTCCTT-3', SEQ ID NO:17 and 5'-TCCCIGCAIAICCAGTAGAARTAGATCTT-3', SEQ ID NO:18. A major PCR product of the expected 1.1 kb size was purified by agarose electrophoresis and used as template in a second PCR amplification reaction.

An aliquot of the RT-PCR product was blunt-

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ended, ligated into a modified Litmus 29 vector and used to transform XL10 competent E. coli. Approximately 120 bacterial colonies were screened for the presence of a fulllength insert by direct PCR using vector primers and Taq polymerase. Plasmids were purified from 25 positive colonies and mapped by digestion with Bam HI. Representative plasmids from each digestion pattern were partially sequenced. Five out of 25 clones contained non-specific amplification products and 20 contained identical inserts similar to human (h)-mox1. One of the latter clones was fully sequenced and found to be 83% identical to h-mox1 over 1060 nucleotides. A 1.1 kb probe was generated by PCR amplification of the insert of a rat mox1 clone with the degenerate primers described above and used to hybridize to a Northern blot of rat vascular smooth muscle cell RNA. A single band, migrating between 28S rRNA and 18S rRNA, indicated the presence of a message with a size compatible to that of human mox-1 (2.6 kb).

To obtain full-length rat mox1, 3' and 5' rapid amplification of cDNA ends (RACE) reactions were performed as describe above, using the gene-specific primers TTGGCACAGTCAGTGAGGATGTCTTC-3', SEQ ID NO:19 5'-CTGTTGGCTTCTACTGTAGCGTTCAAAGTT-3', and SEQ ID NO:20 for 3' and 5' RACE, respectively. Single major 1.5 kb and 850 bp products were obtained for 3' and 5' RACE, These products were purified by agarose gel respectively. eletrophoresis and reamplified with Taq polymerase. products were cloned into the pCR 2.1 vector and used to transform electrocompetent XL1 blue E. coli. The RACE products were sequenced and new terminal primers were designed: 5'sense TTCTGAGTAGGTGTCATAAAGAC-3' ID NO:43), and antisense TTTTCCGTCAAAATTATAACTTTTTATTTTCTTTTATA ACACAT-3' (SEQ ID NO:44). PCR amplification of rat VSMC cDNA was performed using these primers.

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A single 2.6 kb product was obtained, ligated into pCR 2.1 and used to transform electrocompetent XL1 blue E. coli. The insert was sequenced with 12 sense and 14 antisense primers. Its length is 2577 bp (including primer sequences), comprising a 1692 bp open reading frame, 127 bp 5' and 758 bp 3' untranslated regions. The presence of six in-frame stop codons in the 5' untranslated region suggests that the full length coding region has been obtained. Consensus polyadenylation sequences are present at nucleotides 2201 and 2550. Conceptual translation yields a 563 amino acid peptide, one residue shorter than the human deduced sequence. This new amino acid sequence is more similar to human mox1 SEQ ID NO:3 (82% identity) than to mouse gp91phox SEQ ID NO:38 (55% identity), suggesting that it is indeed rat mox1 (SEQ ID NO:21). This rat (r) homolog of p65mox protein is called r-p65mox or p65mox/rat.pep and is shown as SEQ ID NO:21. nucleotide sequence encoding for r-p65mox is shown as SEQ ID NO:22.

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EXAMPLE 13

Expression of rat (r)-p65mox mRNA in Vascular Smooth Muscle and Induction by Angiotensin II, Platelet-Derived Growth Factor (PDGF), and Phorbol Myristic Acid (PMA)

Using the partial cDNA clone from rat, we have examined cultured rat aortic smooth muscle cells for expression of message for r-p65mox. We have observed the mRNA for r-p65mox in these cells. It has been previously reported (Griendling et al. (1994) Circ. Res. 74, 1141-1148; Fukui et al. (1997) Circ. Res. 80, 45-51; Ushio-Fukai et al. (1996) J. Biol. Chem. 271, 23317-23321) that in vitro or in vivo treatment with angiotensin II (AII) is a growth stimulus for vascular smooth muscle cells, and that AII induces increased superoxide generation in these cells. Platelet-derived growth factor (PDGF) and PMA are proliferative signals for vascular smooth muscle cells. We observed that the mRNA for r-p65mox was induced approximately 2-3 fold by angiotensin II (100 nM),

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corresponding to the increased level of superoxide generation. Thus, the increased superoxide generation in these cells correlates with increased expression of the mRNA for this enzyme. The mRNA for r-p65mox also increased 2 or more fold in response to the growth stimulus PDGF (20 ng/ml), and 2-3 fold in response to PMA. Quantitation by densitometry revealed that rat mox1 message was induced nearly 4-fold at the 6 and 12 hour time points in response to PDGF, and about 2-fold at the 12 hour time point in response to AII. 28S RNA was used as a control for RNA recovery.

EXAMPLE 14

Antibodies to Fragments of Human (h)-p65mox (SEQ ID NO:2)

Polyclonal antibodies were raised in rabbits against the C-terminal half of h-p65mox (residues 233 through 564, SEQ ID NO:23) which is predicted to fold into a cytosolic domain containing FAD and the NADPH or NADH binding site. This domain was expressed in E. coli as an N-terminal GST-fusion protein and was purified on glutathione agarose by standard methods. Two antipeptide antibodies were also made against h-p65mox (residues 243-256, referred to as Peptide A, SEQ ID NO:24) and h-65mox (residues 538-551, referred to as Peptide B, SEQ ID NO:25). Peptides were conjugated to keyhole limpet hemocyanin (KLH) using glutaraldehyde.

Antigens were injected into different rabbits initially in complete Freund's adjuvant, and were boosted 4 times with antigen in incomplete Freund's adjuvant at intervals of every three weeks. Approximately 0.5 mg to 1 mg of peptide was administered at each injection. Blood was drawn 1 week after each boost and a terminal bleed was carried out 2 weeks after the final boost. Antibodies to Peptide A and Peptide B were affinity purified by column chromatography through peptide A or peptide B conjugated to Affigel 15 (Bio-Rad, Richmond, CA). 10 mg of peptide was covalently crosslinked to 2 ml of Affigel 15 resin and the gel was washed with 20 ml of binding buffer (20 mM Hepes/NaOH, pH 7.0, 200 mM NaCl,

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and 0.5 % Triton X-100). The remaining functional Nhydrosuccinimide was blocked with 100 µl of 1 ethanolamine. After washing with 20 ml of binding buffer, 5 ml of the antiserum was incubated with the pep A-conjugated Affigel 15 resin overnight at 4°C. Unbound protein was washed away with 20 ml of binding buffer. Elution of the antibodies from the gel was performed with 6 ml of elution buffer (100 mM glycine/HCl, pH 2.5, 200 mM NaCl, and 0.5% Triton X-100). The eluate was then neutralized by adding 0.9 ml of 1 M Tris/HCl, pH 8.0. The GST-fusion form of truncated p65mox1 protein (residues 233-566, SEQ ID NO:23) was expressed in E. coli. Samples (20 µg each) were run on 12 % SDS-PAGE either before or 1 or 4 hours after induction with 100 μM IPTG (isopropyl β-thiogalactoside).

The extracted proteins were subjected to immunoprobing with affinity purified antiserum to peptide A at a 1:1000 dilution. The detection of antigens was performed using an enhanced chemiluminescence kit (Amersham, Buckinghamshire, UK). The affinity purified antibody to mox1 (243-256, SEQ ID NO:24) was used at a dilution of 1:1000 in a Western blot in which a total of 10 µg of protein was added to each lane. The major band observed at 4 hours after IPTG induction corresponded to the size of the GST-mox1 expressed in bacteria containing the pGEX-2T vector encoding the GST-mox1 fusion protein.

Example 15

Presence of an NAD(P)H Oxidase in Ras-Transformed Fibroblasts

A superoxide-generating NADPH oxidase activity was detected in homogenates from NIH 3T3 cells, and this activity increased about 10-15 fold in Ras-transformed NIH 3T3 cells (Table 6). To establish the stable Ras-transformed cell lines, the DNA for human Ras encoding an activating mutation at amino acid number 12 (Valine, referred to as V12-Ras) was subcloned into BamH1 and EcoR1 sites of pCDNA3 vector

which has a neomycin resistance gene. V12-Ras in pCDNA3 and empty vector were transfected into NIH 3T3 cells using Lipofectamine Plus (Gibco). 2 X 10⁶ cells were maintained with DMEM containing 10% calf serum and transfected with 1 mg of DNA. After 2-days, cells were split and selected with the same medium but containing 1 mg/ml neomycin. Colonies surviving in selection media for 10 to 14 days were subcultured and characterized by immunoblot analysis using antibody against human H-Ras.

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The expression of Ras in cells transfected with pcDNA-3 vector alone or in three cell lines transfected with V12-Ras in the same vector was analyzed on a Western blot. The three cell lines were named V12-Ras-7, V12-Ras-4, and V12-Ras-8. The expression of V12-Ras varied widely among the three cell lines tested. The V12-Ras-4 cell line expressed the highest level of Ras followed by the V12-Ras-8 cell line. The V12-Ras-7 cell line expressed the lowest level of Ras.

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Lysates from each of these lines were then prepared and tested for their ability to generate superoxide. For each cell line, cells were washed with cold HANKS' balanced salt solution (HBSS), collected by centrifugation, kept on dry-ice for more than 30 min, and disrupted by suspending in low salt buffer (LSB; 50 mM Tris/HCl, pH 7.5, 1 mM PMSF, and protease cocktail from Sigma) and passing through a syringe needle (18 gauge) ten times. Cell lysates were frozen in dry-ice immediately after determining the protein concentration.

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Table 6 shows superoxide generation in the transfected cells measured using the lucigenin luminescence assay. For the assay, 5 μg of cell lysates were incubated with the reaction mixture containing 10 μM lucigenin (luminescent probe) and 100 μM NADPH (substrate) in the presence or absence of 100 μM arachidonate in the absence or presence of 100 U of superoxide dismutase (SOD) or 1 μM diphenyleneiodonium (DPI). Luminescence of the reaction mixture was monitored for 0.5 second by LumiCounter (Packard) for four times at 3 second intervals. RLU in Table 5

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refers to relative luminescence units.

As shown in Table 6, the luminescence was partially inhibited by superoxide dismutase indicating that the signal was due at least in part to the generation of superoxide. DPI, a known inhibitor of both neutrophil and non-neutrophil NADPH oxidase activities, completely inhibited activity. The generation of superoxide correlated with the expression of Ras in the three cell lines. Thus, oncogenic Ras appears to induce an NADPH-dependent superoxide generating activity that is similar to the activity catalyzed by p65mox1.

Table 6

		RLU/5 µg pr	RLU/5 µg protein			
		no additions	plus SOD	plus DPI		
15	Vector Control (1)	465	154	48		
	V12-Ras-7 (2)	1680	578	39		
	V12-Ras-4 (3)	5975	2128	36		
	V12-Ras-8 (4)	4883	2000	35		

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EXAMPLE 16

Molecular Cloning of Another Rat mox1 cDNA Called Rat mox1B

A rat cDNA library was screened in an effort to identify new rat mox sequences. The library was constructed in a ZAP express lambda phage vector (Stratagene, La Jolla, CA) using RNA isolated from rat vascular smooth muscle cells which had been exposed to 100 nM angiotensin II for 4 hours. The library was screened using standard blot hybridization techniques with the rat mox1 probe described previously. Fifteen individual clones were obtained that were characterized by PCR and restriction mapping. Two different types of clones were thus identified and representatives of each type were sequenced. A clone of the first type (representative of 13) was found to be similar to the previously identified rat mox1 and

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was thus named rat mox1B. Clones of the second type (representative of 2) were incomplete rat mox sequences.

The length of the rat mox1B nucleotide sequence is 2619 bp and is listed as SEQ ID NO:41. The single longest 1497 bp open reading frame encompasses nucleotides 362 to 1858. The presence of two in-frame stop codons in the 5' untranslated region at nucleotides 74 and 257 indicates that the full-length coding region has been isolated. Two putative polyadenylation sites are present at positions 2243 and 2592. Alignment of the rat mox1 nucleotide sequence (SEQ ID NO:22) and the rat mox1B nucleotide sequence (SEQ ID NO:41) shows that the two nucleotides sequences are identical except at their 5' ends, suggesting that they may represent two alternatively spliced messages from the same gene. Sequence identity starts at nucleotides 269 and 311, for rat mox1 and rat mox1B, respectively.

Conceptual translation of the rat mox1B nucleotide sequence (SEQ ID NO:41) yields a 499 amino acid sequence with a predicted molecular weight of 58 kDa. This amino acid sequence for rat mox1B protein is shown in SEQ ID NO:42. Alignment of the deduced amino acid sequences for rat mox1 (SEQ ID NO:21) and rat mox1B (SEQ ID NO:42) indicates that rat mox1B is identical to rat mox1A, except for a missing stretch of 64 residues at the N-terminus. Therefore, rat mox1B appears to be a splicing variant derived from the same gene as rat mox1.

EXAMPLE 17

Sequence Analysis and Cloning of the Human Mox2 cDNA (SEQ ID NO:3) Encoding for Production of the Human Mox2 Protein (SEQ ID NO:4)

Note that the mox2 protein as described herein, was described in U.S. Provisional Application Serial No. 60/149,332 as mox3.

A blast search was carried out using the sequence of mox1 as a query sequence. The sequence identified by this

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search was a sequence present in the GenBank database that contains regions of homology with mox1 and gp91phox. The GenBank sequence located in the search was a 90.6 kb sequenced region of human chromosome 6 (6q25.1-26) that was reported as a GenBank direct submission dated February 9, 1999 and given the Accession No. AL031773. Sequencing was carried out as part of the human genome sequencing project by S. Palmer, at Sanger Centre, in Hinxton, Cambridgeshire, UK. The GenBank sequence was reported as being similar to "Cytochrome B" and was not reported as having any homology or relation to a mox protein. The sequence contained a theoretical amino acid sequence that was derived by computer using an algorithm that predicted intron/exon boundaries and coding regions. This predicted region contained a 545 amino acid sequence that was 56% identical to mox1 and 58% identical to gp91phox.

In the present invention, based on the GenBank genomic sequence and the homologies described above, several specific primers were designed and used to determine the tissue expression patterns of a novel mox protein, mox2, using Human Multiple Tissue PCR Panels (Clontech, Palo Alto, CA). The primers were as follows: Primer 1: 5'-CCTGACAGATGTATTTCACTACCCAG-3' (SEQ ID NO:49); Primer 2: 5'-GGATCGGAGTCACTCCCTTCGCTG-3' (SEQ ID NO:50); Primer 3: 5'-CTAGAAGCTCTCCTTGTTGTAATAGA-3' (SEQ ID NO:51); Primer 4: 5'-ATGAACACCTCTGGGGTCAGCTGA-3' (SEQ ID NO:52). It was determined that mox2 is expressed primarily in fetal tissues, with highest expression in fetal kidney, with expression also seen in fetal liver, fetal lung, fetal brain, fetal spleen and fetal thymus. Among 16 adult tissues tested, mox2 expression was seen in brain, kidney, colon and lung, although levels of expression appeared to be very low.

Additionally, the 5' RACE (RACE = Rapid Amplification of cDNA Ends) and 3' RACE techniques were used to complete the sequence of the 5' and 3' regions of mox2.

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(5' RACE kit and 3' RACE kit were from Clontech, Palo Alto, CA and are more fully described in Frohman et al. (1988) Proc. Natl. Acad. Sci. USA 85, 8998-9002. The 5' RACE and 3'-RACE techniques were carried out using a human fetal kidney library (Marathon-Ready cDNA library, Cat. #7423-1), using the following specific primers: 5'-RACE: Primer 4: 5'-ATGAACACCTCTGGGGTCAGCTGA-3' (SEQ ID NO:53); Primer 5: 5'-GTCCTCTGCAGCATTGTTCCTCTTA-3' (SEQ ID NO:54); 3'-RACE: Primer 1: CCTGACAGATGTATTTCACTACCCAG-3' (SEQ ID NO:55); Primer 2: 5'-GGATCGGAGTCACTCCCTTCGCTG-3' (SEQ ID NO:56). The RACE procedures were successful in completing the 5' sequence and in confirming the 3' sequence. The complete coding sequence of mox2 is shown in SEQ ID NO:2, while the predicted amino acid sequence of mox2 is shown in SEQ ID NO:4.

In comparing the sequences of the present invention to the predicted coding regions of the GenBank sequence, the GenBank sequence did not contain a start codon, appeared to be missing approximately 45 base pairs at the N-terminus, and contained one other major difference in the predicted coding region which could have been due to inaccurate computer prediction of intron/exon boundaries.

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EXAMPLE 18

Sequence Analysis and Partial Cloning of the Human Duox2 cDNA (SEQ ID NO:47) Encoding for Production of the Human Duox2 Protein (SEQ ID NO:48)

A partial cDNA clone of duox2 was obtained as follows.

A 535-base portion of an expressed sequence tag (EST zc92h03.rl; Genbank accession no. W52750) from human pancreatic islet was identified using the human gp91phox amino-acid sequence as a query in a Blast search. The bacterial strain #595758 containing the EST sequence zc92h03.rl in the pBluescript SK-vector was purchased from ATCC (Rockville, MD). The DNA inserted into the pBluescript SK-vector was

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further sequenced using T7 and T3 vector promoters as well as sequence specific internal primers. The EST encoded 440 amino acids showing a 24.4% identity to gp91phox, including a stop codon corresponding to the C-terminus of gp91phox. 5'-RACE was carried out using mRNA obtained from human colon carcinoma cells (CaCo2) and the Marathon cDNA Amplification Kit (ClonTech, Palo Alto). The following genespecific primers were used for this procedure: 5'-GAAGTGGTGGGAGGCGAAGACATA-3' (SEQ ID NO:26) and 5'-CCTGTCATACCTGGGACGGTCTGG-3' (SEQ ID NO:27).

The results of the 5'-RACE yielded an additional 2 kilobase of sequenced DNA but this region did not contain the start codon. To complete the sequence of the 5' and 3' regions of duox2, 5'- RACE and 3'-RACE were carried out using a human adult pancreas mRNA (Clontech, Palo Alto, CA) with the kit of 5' RACE System for Rapid Amplification of cDNA Ends version 2.0 (Gibco BRL, Gaithersburg, MD). PCR done using the following specific primers resulted in a total predicted amino acid sequence of about 1000 residues: 5'-RACE: Primer 5'-GAGCACAGTGAGATGCCTGTTCAG-3' (SEQ ID 3: NO:28); Primer 4: 5'-GGAAGGCAGCAGAGCAATGATG-3' (SEQ ID NO:29) PCR); nested 3'-RACE Primer 5'-ACATCTGCGAGCGCACTTCCAGA-3' (SEQ ID NO:30) Primer 6: 5'-AGCTCGTCAACAGGCAGGACCGAGC-3' (SEQ ID NO:31) (for nested PCR).

EXAMPLE 19

Sequence Analysis and Cloning of the Human Duox1 cDNA (SEQ ID NO:45) Encoding for Production of the Human Duox1 Protein (SEQ ID NO:46)

A cDNA clone of duox1 was obtained as follows. A homologous 357-base portion of an expressed sequence tag (EST nr80d12.s1; Genbank accession no. AA641653) from an invasive human prostate was identified by using the partial

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duox2 predicted amino-acid sequence described above as a query in a Blast search. The bacterial strain #1441736 containing the EST sequence nr80d12.s1 in the pBluescript SKvector was purchased from ATCC (Rockville, MD). The DNA inserted into the pBluescript SK-vector was further sequenced using T7 and T3 vector promoters as well as sequence specific internal primers. The EST insert encoded 673 amino acids with no start or stop codons present. Northern Blot analysis of duox1 indicated the gene was about 5.5 kilobase pairs. complete the sequence of 5' and 3' regions of duox1, 5' RACE and 3'-RACE were carried out using a human adult lung mRNA (Clontech, Palo Alto, CA) with the kit of 5' RACE System for Rapid Amplification of cDNA Ends version 2.0 (Gibco BRL, Gaithersburg, MD). The RACE procedure was carried out using the following specific primers: 5'-RACE: Primer 5: 5'-GCAGTGCATCCACATCTTCAGCAC-3'(SEQ ID NO:32): Primer 5'-GAGAGCTCTGGAGACACTTGAGTTC-3' (SEQ ID NO:33) (for nested PCR); 3'-RACE Primer 7: 5'-CATGTTCTCTCTGGCTGACAAG-3' (SEQ ID Primer 8: 5'-CACAATAGCGAGCTCCGCTTCACGC-3' (SEQ ID NO:35) (for nested PCR). RACE procedures were successful in completing the 5' sequence and the 3' sequence of duox1. The open reading frame is approximately 4563 base pairs.

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EXAMPLE 20

Tissue Expression of Duox1 and Duox2

Based on the duox1 sequence data, several specific primers designed were (Primer 1a: 5'-GCAGGACATCAACCCTGCACTCTC-3' (SEQ ID NO:36); Primer 5'-AATGACACTGTACTGGAGGCCACAG-3' 2a: (SEQ ID NO:57); Primer 3a: 5'-CTGCCATCTACCACACGGATCTGC-3' (SEQ ID NO:58); Primer 4a: 5'-CTTGCCATTCCAAAGCTTCCATGC-3' (SEQ ID NO:59) and used these to determine the tissue expression patterns of duox1 using Human Multiple Tissue PCR Panels

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(Clontech, Palo Alto, CA). It was determined that duox1 is expressed primarily in lung, testis, placenta, prostate, pancreas, fetal heart, fetal kidney, fetal liver, fetal lung, fetal skeletal muscle and thymus, with highest expression in adult and fetal lung. Among 16 adult tissues and 8 fetal tissues tested, duox1 expression in brain, heart, kidney, colon, ovary, thymus, fetal brain and fetal spleen appeared to be low.

Two duox2 specific primers were also used to determine the tissue expression patterns of duox2 using Human Multiple Tissue PCR (polymerase chain reaction) Panels (Clontech, Palo Alto, CA). (Primer 1b: 5'-GTACAAGTCAGGACAGTGGGTGCG-3' (SEQ ID NO:60); Primer 2b: 5'-TGGATGATGTCAGCCAGCCACTCA-3' (SEQ ID NO:61)). Duox2 is expressed primarily in lung, pancreas, placenta, colon, prostate, testis and fetal tissues, with highest expression in adult lung and fetal tissues. Among 16 adult tissues and 8 fetal tissues tested, duox2 expression in brain, heart, kidney, liver, skeletal muscle, thymus and fetal brain appeared to be low.

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EXAMPLE 21

Role of Duox1 and Duox2 in Collagen Crosslinking

To investigate a possible role for the human duox1 and duox2, the model organism *Caenorhabditis elegans* and a new reverse genetic tool, RNA interference (RNAi), were used to "knock out" the homologues of duox in this organism (Fire et al. (1998) *Nature* 391, 806-811). This technique involved injection of double stranded RNA encoding a segment of Ceduox1 or Ceduox2 into gonads of *C. elegans* N2 hermaphrodites. Injected worms were then allowed to lay eggs, and the harvested eggs were allowed to develop and the F1 progeny were scored for phenotypes. This procedure has been documented to "knock-out" the expression of the gene of interest (Fire et al. (1998) *Nature* 391, 806-811).

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In the case of Ce-duox1 and Ce-duox2, the knockout animals resulted in a complex phenotype including

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worms with large superficial blisters, short or "dumpy" worms, worms with locomotion disorders, and worms with retained eggs and/or larvae. Because of the high identity between Ceduox1 and Ce-duox2, three different RNA constructs were predicted to knock out either both genes or Ce-duox2 alone. In all cases, essentially the same group of phenotypes was obtained. Most or all of these phenotypes had been described previously in C. elegans mutated in the collagen biosynthetic pathway. C. elegans has an extracellular structure known as the cuticle, a complex sheath composed largely of cross-linked collagen, which functions as the exoskeleton of the nematode. Cross-linking of collagen in nematodes occurs in part by cross-linking tyrosine residues, and peroxidases such as sea urchin ovoperoxidase and human myeloperoxidase have previously been shown to be capable of carrying out this reaction.

Based upon the similarities of the phenotypes obtained, the Ce-duox1/2 knockout worms were examined for the presence of dityrosine linkages, using an HPLC methodology (Andersen, S.O. (1966) Acta Physiol. Scand. 66, Suppl. 263-265; Abdelrahim et al. (1997) J. Chromatogr. B Biomed. Sci. Appl. 696, 175-182). It was determined that dityrosine linkages, while easily detected in the wild type worms, were almost completely lacking in the knockout worms. Thus, an inability to catalyze dityrosine cross-linking accounts for the phenotype of C. elegans failing to express Ce-duox1/2. These data support the concept that the duox enzymes in higher organisms can probably function in a similar manner to modulate the extracellular milieu, possibly the extracellular matrix and/or the basement membrane.

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All patents, publications and abstracts cited above are incorporated herein by reference in their entirety. It should be understood that the foregoing relates only to preferred embodiments of the present invention and that numerous modifications or alterations may be made therein without

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departing from the spirit and the scope of the present invention as defined in the following claims.

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CLAIMS

- 1. A protein capable of stimulating superoxide production, wherein the protein comprises mox or duox, a fragment thereof or a conservative substitution thereof.
- 2. The protein of Claim 1, wherein the protein, the fragment thereof, or the conservative substitution thereof comprises the amino acid sequence of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:21, SEQ ID NO:42, SEQ ID NO:46, or SEQ ID NO:48, a fragment thereof, or a conservative substitution thereof.
- 3. A nucleotide sequence encoding for the protein, the fragment thereof or the conservative substitution thereof as recited in Claim 1.
- 4. The nucleotide sequence of Claim 3, wherein the nucleotide sequence comprises SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:22, SEQ ID NO:41, SEQ ID NO:45, or SEQ ID NO:47, a fragment thereof, or a conservative substitution thereof.
- 5. A vector, wherein the vector comprises a nucleotide sequence encoding for the protein, the fragment thereof or the conservative substitution thereof, as recited in Claim 1.
 - 6. The vector of Claim 5 wherein the nucleotide sequence comprises SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:22, SEQ ID NO:41, SEQ ID NO:45, or SEQ ID NO:47, a fragment thereof, or a conservative substitution thereof.
 - 7. A cell containing the vector of Claim 5.
 - 8. A cell containing the vector of Claim 6.

- 9. An antibody, wherein the antibody is capable of binding to the protein, the fragment thereof, or the conservative substitution thereof, as recited in Claim 1.
- 10. The antibody of Claim 9, wherein the protein, the fragment thereof, or the conservative substitution thereof, has the amino acid sequence of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:21, SEQ ID NO:42, SEQ ID NO:46, or SEQ ID NO:48, a fragment thereof, or a conservative substitution thereof.

11. A method of stimulating superoxide formation comprising administration, in vitro or in vivo, of a composition comprising the protein, the fragment thereof, or the conservative substitution thereof of Claim 1 in a pharmaceutically acceptable carrier.

- 12. The method of Claim 11, wherein the protein, the fragment thereof, or the conservative substitution thereof, comprises the amino acid sequence of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:45, SEQ ID NO:46, or SEQ ID NO:48, a fragment thereof, or a conservative substitution thereof.
- 13. A method of stimulating superoxide formation comprising administration, in vitro or in vivo, of a composition comprising the vector of Claim 5 in a pharmaceutically acceptable carrier.
- 14. A method of stimulating superoxide formation comprising administration, in vitro or in vivo, of a composition comprising the vector of Claim 6 in a pharmaceutically acceptable carrier.
- 15. A method for determining the activity of a drug comprising measuring the activity of the protein, the fragment thereof or the conservative substitution thereof, as recited in

Claim 1, to stimulate superoxide production following administration of the drug.

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16. The method of Claim 15, wherein the protein, the fragment thereof or the conservative substitution thereof comprises the amino acid sequence of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:45, or SEQ ID NO:48, a fragment thereof, or a conservative substitution thereof.

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         MLSQKLIPTKDRNPVKRFAMNISYFFLENW cytb558/arabidopsis.pep
---NLAGLRKKSSIRKISTSLSYYFEDNW cytb558.rice.pep
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       LSIFAILVWLGLNVFLFV-WYYRVYDIPPK GDALDDX.DIMMAN
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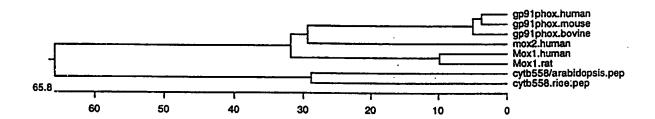
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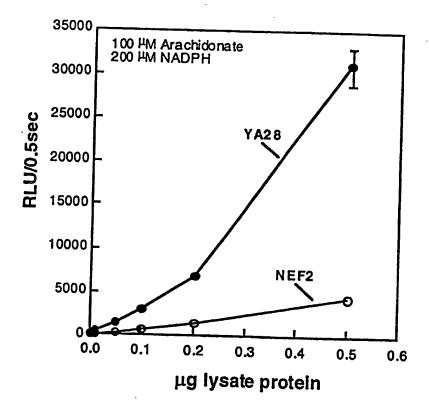
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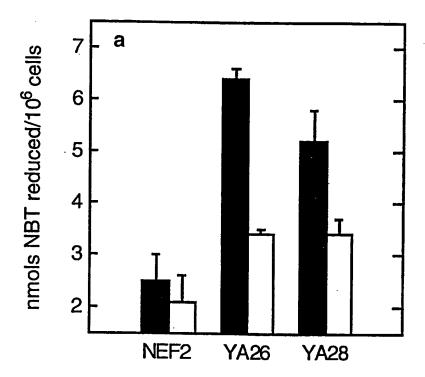
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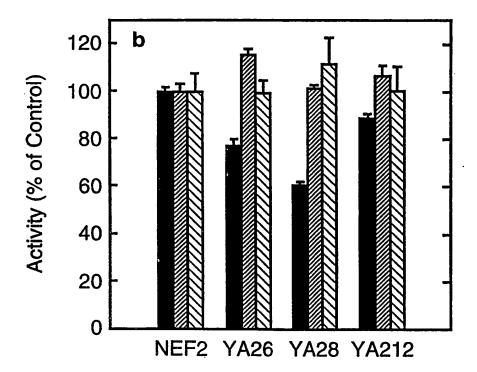
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-++	+	+ ~~	202	~~>		+ ~ ~								_ 4		400
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acc	act	gaa	ttg	cta	agg	aca	ata	qca	aac	qtc	acc	aat	cta	ata	atc	643
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C24	acc	tee	tat		++~	++~	+~~	+==	30"	~~	cat			-+-		720
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215 220 225

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				ccc Pro				1363

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Trp Tyr Glu Glu Glu Glu Ser Phe His Tyr Thr Arg Val Ile Leu Gly 35 40 45

Ser Thr Leu Ala Trp Ala Arg Ala Ser Ala Leu Cys Leu Asn Phe Asn 50 55 60

Cys Met Leu Ile Leu Ile Pro Val Ser Arg Asn Leu Ile Ser Phe Ile 65 70 75 80

Arg Gly Thr Ser Ile Cys Cys Arg Gly Pro Trp Arg Arg Gln Leu Asp 85 90 95

Lys Asn Leu Arg Phe His Lys Leu Val Ala Tyr Gly Ile Ala Val Asn 100 105 110

Ala Thr Ile His Ile Val Ala His Phe Phe Asn Leu Glu Arg Tyr His 115 120 125

Trp Ser Gln Ser Glu Glu Ala Gln Gly Leu Leu Ala Ala Leu Ser Lys 130 135 140

Leu Gly Asn Thr Pro Asn Glu Ser Tyr Leu Asn Pro Val Arg Thr Phe 145 150 155 160

Pro Thr Asn Thr Thr Glu Leu Leu Arg Thr Ile Ala Gly Val Thr 165 170 175

Gly Leu Val Ile Ser Leu Ala Leu Val Leu Ile Met Thr Ser Ser Thr 180 185 190

Glu Phe Ile Arg Gln Ala Ser Tyr Glu Leu Phe Trp Tyr Thr His His 195 200 205

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- Val Pro Gln Phe Ser Gly Lys Glu Pro Ser Ala Trp Lys Trp Ile Leu 260 265 270
- Gly Pro Val Val Leu Tyr Ala Cys Glu Arg Ile Ile Arg Phe Trp Arg 275 280 285
- Phe Gln Gln Glu Val Val Ile Thr Lys Val Val Ser His Pro Ser Gly 290 295 300
- Val Leu Glu Leu His Met Lys Lys Arg Gly Phe Lys Met Ala Pro Gly 305 310 315 320
- Gln Tyr Ile Leu Val Gln Cys Pro Ala Ile Ser Ser Leu Glu Trp His 325 330 335
- Pro Phe Thr Leu Thr Ser Ala Pro Gln Glu Asp Phe Phe Ser Val His 340 345 350
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- Val Asp Gly Pro Phe Gly Thr Ala Leu Thr Asp Val Phe His Tyr Pro 385 390 395 400
- Val Cys Val Cys Val Ala Ala Gly Ile Gly Val Thr Pro Phe Ala Ala 405 410 415
- Leu Leu Lys Ser Ile Trp Tyr Lys Cys Ser Glu Ala Gln Thr Pro Leu
 420 425 430
- Lys Leu Ser Lys Val Tyr Phe Tyr Trp Ile Cys Arg Asp Ala Arg Ala 435 440 445
- Phe Glu Trp Phe Ala Asp Leu Leu Leu Ser Leu Glu Thr Arg Met Ser 450 455 460

Glu Gln Gly Lys Thr His Phe Leu Ser Tyr His Ile Phe Leu Thr Gly 470 Trp Asp Glu Asn Gln Ala Leu His Ile Ala Leu His Trp Asp Glu Asn 485 490 Thr Asp Val Ile Thr Gly Leu Lys Gln Lys Thr Phe Tyr Gly Arg Pro 500 505 Asn Trp Asn Asn Glu Phe Lys Gln Ile Ala Tyr Asn His Pro Ser Ser 515 520 Ser Ile Gly Val Phe Phe Cys Gly Pro Lys Ala Leu Ser Arg Thr Leu 535 Gln Lys Met Cys His Leu Tyr Ser Ser Ala Asp Pro Arg Gly Val His 545 550 555 Phe Tyr Tyr Asn Lys Glu Ser Phe 565 <210> 5 <211> 21 <212> DNA <213> Artificial Sequence <220> <223> Description of Artificial Sequence: primer <400> 5 aacaagcgtg gcttcagcat g 21 <210> 6 <211> 18 <212> DNA <213> Artificial Sequence <220> <223> Description of Artificial Sequence: primer <400> 6 agcaatattg ttggtcat

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Leu Ala Leu Ala Arg Ala Pro Ala Ala Cys Leu Asn Phe Asn Cys Met 50 55 60

Leu Ile Leu Leu Pro Val Cys Arg Asn Leu Leu Ser Phe Leu Arg Gly
65 70 75 80

Ser Ser Ala Cys Cys Ser Thr Arg Val Arg Arg Gln Leu Asp Arg Asn 85 90 95

Leu Thr Phe His Lys Met Val Ala Trp Met Ile Ala Leu His Ser Ala
100 105 110

Ile His Thr Ile Ala His Leu Phe Asn Val Glu Trp Cys Val Asn Ala 115 120 125

Arg Val Asn Asn Ser Asp Pro Tyr Ser Val Ala Leu Ser Glu Leu Gly
130 135 140

Asp Arg Gln Asn Glu Ser Tyr Leu Asn Phe Ala Arg Lys Arg Ile Lys 145 150 155 160

Asn Pro Glu Gly Gly Leu Tyr Leu Ala Val Thr Leu Leu Ala Gly Ile 165 170 175

Thr Gly Val Val Ile Thr Leu Cys Leu Ile Leu Ile Ile Thr Ser Ser 180 185 190

- Thr Lys Thr Ile Arg Arg Ser Tyr Phe Glu Val Phe Trp Tyr Thr His 195 200 205
- His Leu Phe Val Ile Phe Phe Ile Gly Leu Ala Ile His Gly Ala Glu 210 215 220
- Arg Ile Val Arg Gly Gln Thr Ala Glu Ser Leu Ala Val His Asn Ile 225 230 235 240
- Thr Val Cys Glu Gln Lys Ile Ser Glu Trp Gly Lys Ile Lys Glu Cys 245 250 255
- Pro Ile Pro Gln Phe Ala Gly Asn Pro Pro Met Thr Trp Lys Trp Ile 260 265 270
- Val Gly Pro Met Phe Leu Tyr Leu Cys Glu Arg Leu Val Arg Phe Trp
 275 280 285
- Arg Ser Gln Gln Lys Val Val Ile Thr Lys Val Val Thr His Pro Phe 290 295 300
- Lys Thr Ile Glu Leu Gln Met Lys Lys Lys Gly Phe Lys Met Glu Val 305 310 315 320
- Gly Gln Tyr Ile Phe Val Lys Cys Pro Lys Val Ser Lys Leu Glu Trp 325 330 335
- His Pro Phe Thr Leu Thr Ser Ala Pro Glu Glu Asp Phe Phe Ser Ile 340 345 350
- His Ile Arg Ile Val Gly Asp Trp Thr Glu Gly Leu Phe Asn Ala Cys 355 360 365
- Gly Cys Asp Lys Gln Glu Phe Gln Asp Ala Trp Lys Leu Pro Lys Ile 370 375 380
- Ala Val Asp Gly Pro Phe Gly Thr Ala Ser Glu Asp Val Phe Ser Tyr 385 390 395 400
- Glu Val Val Met Leu Val Gly Ala Gly Ile Gly Val Thr Pro Phe Ala 405 410 415
- Ser Ile Leu Lys Ser Val Trp Tyr Lys Tyr Cys Asn Asn Ala Thr Asn 420 425 430

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WO 00/28031

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Tyr Glu Lys Ser Asp Lys Tyr Tyr Tyr Thr Arg Glu Ile Leu Gly Thr
35 40 45

Ala Leu Ala Leu Ala Arg Ala Ser Ala Leu Cys Leu Asn Phe Asn Ser 50 55 60

Met Val Ile Leu Ile Pro Val Cys Arg Asn Leu Leu Ser Phe Leu Arg
65 70 75 80

Gly Thr Cys Ser Phe Cys Asn His Thr Leu Arg Lys Pro Leu Asp His 85 90 95

Asn Leu Thr Phe His Lys Leu Val Ala Tyr Met Ile Cys Ile Phe Thr

Ala Ile His Ile Ile Ala His Leu Phe Asn Phe Glu Arg Tyr Ser Arg
115 120 125

Ser Gln Gln Ala Met Asp Gly Ser Leu Ala Ser Val Leu Ser Ser Leu 130 135 140

Phe His Pro Glu Lys Glu Asp Ser Trp Leu Asn Pro Ile Gln Ser Pro 145 150 155 160

Asn Val Thr Val Met Tyr Ala Ala Phe Thr Ser Ile Ala Gly Leu Thr

165 170 175

Gly Val Val Ala Thr Val Ala Leu Val Leu Met Val Thr Ser Ala Met
180 185 190

- Glu Phe Ile Arg Arg Asn Tyr Phe Glu Leu Phe Trp Tyr Thr His His 195 200 205
- Leu Phe Ile Ile Tyr Ile Ile Cys Leu Gly Ile His Gly Leu Gly Gly 210 215 220
- Ile Val Arg Gly Gln Thr Glu Glu Ser Met Ser Glu Ser His Pro Arg 225 230 235 240
- Asn Cys Ser Tyr Ser Phe His Glu Trp Asp Lys Tyr Glu Arg Ser Cys
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- Arg Ser Pro His Phe Val Gly Gln Pro Pro Glu Ser Trp Lys Trp Ile 260 265 270
- Leu Ala Pro Ile Ala Phe Tyr Ile Phe Glu Arg Ile Leu Arg Phe Tyr
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- Gly Gln Tyr Ile Phe Val Asn Cys Pro Ser Ile Ser Phe Leu Glu Trp
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- His Pro Phe Thr Leu Thr Ser Ala Pro Glu Glu Glu Phe Phe Ser Ile 340 345 350
- His Ile Arg Ala Ala Gly Asp Trp Thr Glu Asn Leu Ile Arg Thr Phe 355 360 365
- Glu Gln Gln His Ser Pro Met Pro Arg Ile Glu Val Asp Gly Pro Phe 370 375 380
- Gly Thr Val Ser Glu Asp Val Phe Gln Tyr Glu Val Ala Val Leu Val 385 390 395 400
- Gly Ala Gly Ile Gly Val Thr Pro Phe Ala Ser Phe Leu Lys Ser Ile
 405 410 415
- Trp Tyr Lys Phe Gln Arg Ala His Asn Lys Leu Lys Thr Gln Lys Ile

420

425

430

Tyr Phe Tyr Trp Ile Cys Arg Glu Thr Gly Ala Phe Ala Trp Phe Asn 435 440 445

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Asp Phe Leu Asn Tyr Arg Leu Phe Leu Thr Gly Trp Asp Ser Asn Ile 465 470 475 480

Ala Gly His Ala Ala Leu Asn Phe Asp Arg Ala Thr Asp Val Leu Thr 485 490 495

Gly Leu Lys Gln Lys Thr Ser Phe Gly Arg Pro Met Trp Asp Asn Glu
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Phe Ser Arg Ile Ala Thr Ala His Pro Lys Ser Val Val Gly Val Phe 515 520 525

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Arg Tyr Ser Ser Leu Asp Pro Arg Lys Val Gln Phe Tyr Phe Asn Lys 545 550 555 560

Glu Thr Phe

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Met Trp Asp Asp Arg Asp Ser His Cys Arg Arg Pro Lys Phe Glu Gly
20 25 30

His Pro Pro Glu Ser Trp Lys Trp Ile Leu Ala Pro Val Ile Leu Tyr 35 40 45

Ile Cys Glu Arg Ile Leu Arg Phe Tyr Arg Ser Gln Gln Lys Val Val
50 55 60

Ile Thr Lys Val Val Met His Pro Ser Lys Val Leu Glu Leu Gln Met 65 70 75 80

Asn Lys Arg Gly Phe Ser Met Glu Val Gly Gln Tyr Ile Phe Val Asn 85 90 95

- Cys Pro Ser Ile Ser Leu Leu Glu Trp His Pro Phe Thr Leu Thr Ser 100 105 110
- Ala Pro Glu Glu Asp Phe Phe Ser Ile His Ile Arg Ala Ala Gly Asp 115 120 125
- Trp Thr Glu Asn Leu Ile Arg Ala Phe Glu Gln Gln Tyr Ser Pro Ile 130 135 140
- Pro Arg Ile Glu Val Asp Gly Pro Phe Gly Thr Ala Ser Glu Asp Val 145 150 155 160
- Phe Gln Tyr Glu Val Ala Val Leu Val Gly Ala Gly Ile Gly Val Thr 165 170 175
- Pro Phe Ala Ser Ile Leu Lys Ser Ile Trp Tyr Lys Phe Gln Cys Ala 180 185 190
- Asp His Asn Leu Lys Thr Lys Lys Ile Tyr Phe Tyr Trp Ile Cys Arg
- Glu Thr Gly Ala Phe Ser Trp Phe Asn Asn Leu Leu Thr Ser Leu Glu 210 215 220
- Gln Glu Met Glu Glu Leu Gly Lys Val Gly Phe Leu Asn Tyr Arg Leu 225 230 235 240
- Phe Leu Thr Gly Trp Asp Ser Asn Ile Val Gly His Ala Ala Leu Asn 245 250 255
- Phe Asp Lys Ala Thr Asp Ile Val Thr Gly Leu Lys Gln Lys Thr Ser 260 265 270
- Phe Gly Arg Pro Met Trp Asp Asn Glu Phe Ser Thr Ile Ala Thr Ser 275 280 285
- His Pro Lys Ser Val Val Gly Val Phe Leu Cys Gly Pro Arg Thr Leu 290 295 300
- Ala Lys Ser Leu Arg Lys Cys Cys His Arg Tyr Ser Ser Leu Asp Pro 305 310 315 320
- Arg Lys Val Gln Phe Tyr Phe Asn Lys Glu Asn Phe 325 330

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<213> Homo sapiens
<400> 24
Cys Ala Glu Ser Phe Glu Met Trp Asp Asp Arg Asp Ser His
<210> 25
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Lys Ser Leu Arg Lys Cys Cys His Arg Tyr Ser Ser Leu Asp
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                                      10
<210> 26
<211> 24
<212> DNA
<213> Artificial Sequence
<223> Description of Artificial Sequence: primer
<400> 26
gaagtggtgg gaggcgaaga cata
                                                                     24
<210> 27
<211> 24
<212> DNA
<213> Artificial Sequence
<223> Description of Artificial Sequence: primer
<400> 27
cctgtcatac ctgggacggt ctgg
                                                                     24
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<211> 24
<212> DNA
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WO 00/28031	PCT/US99/26592
<213> Artificial Sequence	
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<223> Description of Artificial Sequence: primer	
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gagcacagtg agatgcctgt tcag	24
	24
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<211> 24 <212> DNA	
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<220>	
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1 Parmer	
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ggaaggcagc agagagcaat gatg	24
<210> 30	
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<212> DNA	
<213> Artificial Sequence	•
<220>	
<223> Description of Artificial Sequence: primer	
4400. 00	
<400> 30	
acatctgcga gcggcacttc caga	24
<210> 31	
<211> 25	
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agctcgtcaa caggcaggac cgagc	
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<210> 32	
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<212> DNA	

WO 00/28031	PCT/US99/26592
<213> Artificial Sequence	
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gcagtgcatc cacatettca gcac	24
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<211> 25	
<212> DNA	
<213> Artificial Sequence	
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gagagetetg gagacaettg agtte	25
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<210> 34	
<211> 22	
<212> DNA	
<213> Artificial Sequence	
<220>	
<223> Description of Artificial Sequence: primer	
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<210> 35	
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<223> Description of Artificial Sequence: primer	
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<211> 24	
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<213> Artificial Sequence

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24

<210> 37

<211> 570

<212> PRT

<213> Bovine

<400> 37

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Val Trp Leu Gly Met Asn Val Phe Leu Phe Val Trp Tyr Tyr Arg Val 20 25 30

Tyr Asp Ile Pro Asp Lys Phe Phe Tyr Thr Arg Lys Leu Leu Gly Ser 35 40 45

Ala Leu Ala Leu Ala Arg Ala Pro Ala Ala Cys Leu Asn Phe Asn Cys
50 55 60

Met Leu Ile Leu Leu Pro Val Cys Arg Asn Leu Leu Ser Phe Leu Arg 65 70 75 80

Gly Ser Ser Ala Cys Cys Ser Thr Arg Ile Arg Arg Gln Leu Asp Arg 85 90 95

Asn Leu Thr Phe His Lys Met Val Ala Trp Met Ile Ala Leu His Thr 100 105 110

Ala Ile His Thr Ile Ala His Leu Phe Asn Val Glu Trp Cys Val Asn 115 120 125

Ala Arg Val Asn Asn Ser Asp Pro Tyr Ser Ile Ala Leu Ser Asp Ile 130 135 140

Gly Asp Lys Pro Asn Glu Thr Tyr Leu Asn Phe Val Arg Gln Arg Ile
145 150 155 160

Lys Asn Pro Glu Gly Gly Leu Tyr Val Ala Val Thr Arg Leu Ala Gly 165 170 175 Ile Thr Gly Val Val Ile Thr Leu Cys Leu Ile Leu Ile Ile Thr Ser 180 185 190

- Ser Thr Lys Thr Ile Arg Arg Ser Tyr Phe Glu Val Phe Trp Tyr Thr 195 200 205
- His His Leu Phe Val Ile Phe Phe Ile Gly Leu Ala Ile His Gly Ala 210 215 220
- Gln Arg Ile Val Arg Gly Gln Thr Ala Glu Ser Leu Leu Lys His Gln 225 230 235 240
- Pro Arg Asn Cys Tyr Gln Asn Ile Ser Gln Trp Gly Lys Ile Glu Asn 245 250 255
- Cys Pro Ile Pro Glu Phe Ser Gly Asn Pro Pro Met Thr Trp Lys Trp 260 265 270
- Ile Val Gly Pro Met Phe Leu Tyr Leu Cys Glu Arg Leu Val Arg Phe 275 280 285
- Trp Arg Ser Gln Gln Lys Val Val Ile Thr Lys Val Val Thr His Pro 290 295 300
- Phe Lys Thr Ile Glu Leu Gln Met Lys Lys Lys Gly Phe Lys Met Glu 305 310 315 320
- Val Gly Gln Tyr Ile Phe Val Lys Cys Pro Val Val Ser Lys Leu Glu 325 330 335
- Trp His Pro Phe Thr Leu Thr Ser Ala Pro Glu Glu Asp Phe Phe Ser 340 345 350
- Ile His Ile Arg Ile Val Gly Asp Trp Thr Glu Gly Leu Phe Lys Ala 355 360 365
- Cys Gly Cys Asp Lys Gln Glu Phe Gln Asp Ala Trp Lys Leu Pro Lys 370 375 380
- Ile Ala Val Asp Gly Pro Phe Gly Thr Ala Ser Glu Asp Val Phe Ser 385 390 395 400
- Tyr Glu Val Val Met Leu Val Gly Ala Gly Ile Gly Val Thr Pro Phe
 405 410 415
- Ala Ser Ile Leu Lys Ser Val Trp Tyr Lys Tyr Cys Asn Lys Ala Pro 420 425 430

Asn Leu Arg Leu Lys Lys Ile Tyr Phe Tyr Trp Leu Cys Arg Asp Thr 435 440 445

His Ala Phe Glu Trp Phe Ala Asp Leu Leu Gln Leu Leu Glu Thr Gln 450 455 460

Met Gln Glu Lys Asn Asn Thr Asp Phe Leu Ser Tyr Asn Ile Cys Leu 465 470 475 480

Thr Gly Trp Asp Glu Ser Gln Ala Ser His Phe Ala Met His His Asp
485 490 495

Glu Glu Lys Asp Val Ile Thr Gly Leu Lys Gln Lys Thr Leu Tyr Gly
500 505 510

Arg Pro Asn Trp Asp Asn Glu Phe Lys Thr Ile Gly Ser Gln His Pro 515 520 525

Asn Thr Arg Ile Gly Val Phe Leu Cys Gly Pro Glu Ala Leu Ala Asp 530 540

Thr Leu Asn Lys Gln Cys Ile Ser Asn Ser Asp Ser Gly Pro Arg Gly 545 550 555 560

Val His Phe Ile Phe Asn Lys Glu Asn Phe 565 570

<210> 38

<211> 570

<212> PRT

<213> murine

<400> 38

Met Gly Asn Trp Ala Val Asn Glu Gly Leu Ser Ile Phe Val Ile Leu
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Val Trp Leu Gly Leu Asn Val Phe Leu Phe Ile Asn Tyr Tyr Lys Val 20 25 30

Tyr Asp Asp Gly Pro Lys Tyr Asn Tyr Thr Arg Lys Leu Leu Gly Ser 35 40 45

Ala Leu Ala Leu Ala Arg Ala Pro Ala Ala Cys Leu Asn Phe Asn Cys
50 55 60

Met Leu Ile Leu Leu Pro Val Cys Arg Asn Leu Leu Ser Phe Leu Arg 65 70 75 80

Gly	Ser	Ser	Ala	Cys 85	Cys	Ser	Thr	Arg	Ile 90		Arg	Gln	Leu	Asp 95	Arg
Asn	Leu	Thr	Phe 100	His	Lys	Met	Val	Ala 105	Trp	Met	Ile	Ala	Leu 110	His	Thr
Ala	Ile	His 115	Thr	Ile	Ala	His	Leu 120	Phe	Asn	Val	Glu	Trp 125	Cys	Val	Asn
Ala	Arg 130	Val	Gly	Ile	Ser	Asp 135	Arg	Tyr	Ser	Ile	Ala 140	Leu	Ser	Asp	Ile
Gly 145	Asp	Asn	Glu	Asn	Glu 150	Glu	Tyr	Leu	Asn	Phe 155	Ala	Arg	Glu	Lys	Ile 160
Lys	Asn	Pro	Glu	Gly 165	Gly	Leu	Tyr	Val	Ala 170	Val	Thr	Arg	Leu	Ala 175	Gly
Ile	Thr	Gly	Ile 180	Val	Ile	Thr	Leu	Cys 185	Leu	Ile	Leu	Ile	Ile 190	Thr	Ser
Ser	Thr	Lys 195	Thr	Ile	Arg	Arg	Ser 200	Tyr	Phe	Glu	Val	Phe 205	Trp	Tyr	Thr
His	His 210	Leu	Phe	Val	Ile	Phe 215	Phe	Ile	GŢĀ	Leu	Ala 220	Ile	His	Gly	Ala
Glu 225	Arg	Ile	Val	Arg	Gly 230	Gln	Thr	Ala	Glu	Ser 235	Leu	Glu	Glu	His	Asn 240
Leu	Asp	Ile	Cys	Ala 245	Asp	Lys	Ile	Glu	Glu 250	Trp	Gly	Lys	Ile	Lys 255	Glu
Cys	Pro	Val	Pro 260	Lys	Phe	Ala	Gly	Asn 265	Pro	Pro	Met	Thr	Trp 270	Lys	Trp
Ile	Val	Gly 275	Pro	Met	Phe	Leu	Tyr 280	Leu	Cys	Glu	Arg	Leu 285	Val	Arg	Phe
Trp	Arg 290	Ser	Gln	Gln	Lys	Val 295	Val	Ile	Thr	Lys	Val 300	Val	Thr	His	Pro
Phe 305	Lys	Thr	Ile.	Glu	Leu 310	Gln	Met	Lys		Lys 315	Gly	Phe	Lys		Glu 320

325

Val Gly Gln Tyr Ile Phe Val Lys Cys Pro Lys Val Ser Lys Leu Glu

330

Trp His Pro Phe Thr Leu Thr Ser Ala Pro Glu Glu Asp Phe Phe Ser 340 345 350

Ile His Ile Arg Ile Val Gly Asp Trp Thr Glu Gly Leu Phe Asn Ala 355 360 365

Cys Gly Cys Asp Lys Gln Glu Phe Gln Asp Ala Trp Lys Leu Pro Lys 370 380

Ile Ala Val Asp Gly Pro Phe Gly Thr Ala Ser Glu Asp Val Phe Ser 385 390 395 400

Tyr Glu Val Val Met Leu Val Gly Ala Gly Ile Gly Val Thr Pro Phe
405 410 415

Ala Ser Ile Leu Lys Ser Val Trp Tyr Lys Tyr Cys Asp Asn Ala Thr 420 425 430

Ser Leu Lys Leu Lys Lys Ile Tyr Phe Tyr Trp Leu Cys Arg Asp Thr 435 440 445

His Ala Phe Glu Trp Phe Ala Asp Leu Leu Gln Leu Leu Glu Thr Gln 450 455 460

Met Gln Glu Arg Asn Asn Ala Asn Phe Leu Ser Tyr Asn Ile Tyr Leu 465 470 475 480

Thr Gly Trp Asp Glu Ser Gln Ala Asn His Phe Ala Val His His Asp 485 490 495

Glu Glu Lys Asp Val Ile Thr Gly Leu Lys Gln Lys Thr Leu Tyr Gly
500 505 510

Arg Pro Asn Trp Asp Asn Glu Phe Lys Thr Ile Ala Ser Glu His Pro 515 520 525

Asn Thr Thr Ile Gly Val Phe Leu Cys Gly Pro Glu Ala Leu Ala Glu 530 535 540

Thr Leu Ser Lys Gln Ser Ile Ser Asn Ser Glu Ser Gly Pro Arg Gly 545 550 555 560

Val His Phe Ile Phe Asn Lys Glu Asn Phe 565 570

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<211> 944

<212> PRT

<213> Arabidopsis sp.

<400> 39

Met Lys Pro Phe Ser Lys Asn Asp Arg Arg Arg Trp Ser Phe Asp Ser 1 5 10 15

Val Ser Ala Gly Lys Thr Ala Val Gly Ser Ala Ser Thr Ser Pro Gly
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Thr Glu Tyr Ser Ile Asn Gly Asp Gln Glu Phe Val Glu Val Thr Ile
35 40 45

Asp Leu Gln Asp Asp Asp Thr Ile Val Leu Arg Ser Val Glu Pro Ala
50 55 60

Thr Ala Ile Asn Val Ile Gly Asp Ile Ser Asp Asp Asn Thr Gly Ile
65 70 75 80

Met Thr Pro Val Ser Ile Ser Arg Ser Pro Thr Met Lys Arg Thr Ser 85 90 95

Ser Asn Arg Phe Arg Gln Phe Ser Gln Glu Leu Lys Ala Glu Ala Val

Ala Lys Ala Lys Gln Leu Ser Gln Glu Leu Lys Arg Phe Ser Trp Ser 115 120 125

Arg Ser Phe Ser Gly Asn Leu Thr Thr Thr Ser Thr Ala Ala Asn Gln
130 135 140

Ser Gly Gly Ala Gly Gly Gly Leu Val Asn Ser Ala Leu Glu Ala Arg 145 150 155 160

Ala Leu Arg Lys Gln Arg Ala Gln Leu Asp Arg Thr Arg Ser Ser Ala 165 170 175

Gln Arg Ala Leu Arg Gly Leu Arg Phe Ile Ser Asn Lys Gln Lys Asn 180 185 190

Val Asp Gly Trp Asn Asp Val Gln Ser Asn Phe Glu Lys Phe Glu Lys
195 200 205

Asn Gly Tyr Ile Tyr Arg Ser Asp Phe Ala Gln Cys Ile Gly Met Lys 210 215 220

Asp Ser Lys Glu Phe Ala Leu Glu Leu Phe Asp Ala Leu Ser Arg Arg

225 230 235 240

Arg Arg Leu Lys Val Glu Lys Ile Asn His Asp Glu Leu Tyr Glu Tyr
245 250 255

Trp Ser Gln Ile Asn Asp Glu Ser Phe Asp Ser Arg Leu Gln Ile Phe 260 265 270

Phe Asp Ile Val Asp Lys Asn Glu Asp Gly Arg Ile Thr Glu Glu Glu 275 280 285

Val Lys Glu Ile Ile Met Leu Ser Ala Ser Ala Asn Lys Leu Ser Arg 290 295 300

Leu Lys Glu Gln Ala Glu Glu Tyr Ala Ala Leu Ile Met Glu Glu Leu 305 310 315 320

Asp Pro Glu Arg Leu Gly Tyr Ile Glu Leu Trp Gln Leu Glu Thr Leu 325 330 335

Leu Leu Gln Lys Asp Thr Tyr Leu Asn Tyr Ser Gln Ala Leu Ser Tyr 340 345 350

Thr Ser Gln Ala Leu Ser Gln Asn Leu Gln Gly Leu Arg Gly Lys Ser 355 360 365 .

Arg Ile His Arg Met Ser Ser Asp Phe Val Tyr Ile Met Gln Glu Asn 370 380

Trp Lys Arg Ile Trp Val Leu Ser Leu Trp Ile Met Ile Met Ile Gly 385 390 395 400

Leu Phe Leu Trp Lys Phe Phe Gln Tyr Lys Gln Lys Asp Ala Phe His
405 410 415

Val Met Gly Tyr Cys Leu Leu Thr Ala Lys Gly Ala Ala Glu Thr Leu
420 425 430

Lys Phe Asn Met Ala Leu Ile Leu Phe Pro Val Cys Arg Asn Thr Ile
435
440
445

Thr Trp Leu Arg Ser Thr Arg Leu Ser Tyr Phe Val Pro Phe Asp Asp 450 455 460

Asn Ile Asn Phe His Lys Thr Ile Ala Gly Ala Ile Val Val Ala Val
465 470 475 480

Ile Leu His Ile Gly Asp His Leu Ala Cys Asp Phe Pro Arg Ile Val

495

485 490

Arg Ala Thr Glu Tyr Asp Tyr Asn Arg Tyr Leu Phe His Tyr Phe Gln 500 505 510

Thr Lys Gln Pro Thr Tyr Phe Asp Leu Val Lys Gly Pro Glu Gly Ile
515 520 525

Thr Gly Ile Leu Met Val Ile Leu Met Ile Ile Ser Phe Thr Leu Ala 530 535 540

Thr Arg Trp Phe Arg Arg Asn Leu Val Lys Leu Pro Lys Pro Phe Asp 545 550 555 560

Arg Leu Thr Gly Phe Asn Ala Phe Trp Tyr Ser His His Leu Phe Val

Ile Val Tyr Ile Leu Leu Ile Leu His Gly Ile Phe Leu Tyr Phe Ala
580 585 590

Lys Pro Trp Tyr Val Arg Thr Thr Trp Met Tyr Leu Ala Val Pro Val 595 600 605

Leu Leu Tyr Gly Gly Glu Arg Thr Leu Arg Tyr Phe Arg Ser Gly Ser 610 620

Tyr Ser Val Arg Leu Leu Lys Val Ala Ile Tyr Pro Gly Asn Val Leu 625 630 635 640

Thr Leu Gln Met Ser Lys Pro Thr Gln Phe Arg Tyr Lys Ser Gly Gln
645 650 655

Tyr Met Phe Val Gln Cys Pro Ala Val Ser Pro Phe Glu Trp His Pro 660 665 670

Phe Ser Ile Thr Ser Ala Pro Glu Asp Asp Tyr Ile Ser Ile His Ile 675 680 685

Arg Gln Leu Gly Asp Trp Thr Gln Glu Leu Lys Arg Val Phe Ser Glu 690 695 700

Val Cys Glu Pro Pro Val Gly Gly Lys Ser Gly Leu Leu Arg Ala Asp 705 710 715 720

Glu Thr Thr Lys Lys Ser Leu Pro Lys Leu Leu Ile Asp Gly Pro Tyr 725 730 735

Gly Ala Pro Ala Gln Asp Tyr Arg Lys Tyr Asp Val Leu Leu Val

740

745

750

Gly Leu Gly Ile Gly Ala Thr Pro Phe Ile Ser Ile Leu Lys Asp Leu 755 760 765

Leu Asn Asn Ile Val Lys Met Glu Glu His Ala Asp Ser Ile Ser Asp 770 780

Phe Ser Arg Ser Ser Glu Tyr Ser Thr Gly Ser Asn Gly Asp Thr Pro
785 790 795 800

Arg Arg Lys Arg Ile Leu Lys Thr Thr Asn Ala Tyr Phe Tyr Trp Val

Thr Arg Glu Gln Gly Ser Phe Asp Trp Phe Lys Gly Val Met Asn Glu 820 825 830

Val Ala Glu Leu Asp Gln Arg Gly Val Ile Glu Met His Asn Tyr Leu 835 840 845

Thr Ser Val Tyr Glu Glu Gly Asp Ala Arg Ser Ala Leu Ile Thr Met 850 855 860

Val Gln Ala Leu Asn His Ala Lys Asn Gly Val Asp Ile Val Ser Gly 865 870 875 880

Thr Arg Val Arg Thr His Phe Ala Arg Pro Asn Trp Lys Lys Val Leu 885 890 895

Thr Lys Leu Ser Ser Lys His Cys Asn Ala Arg Ile Gly Val Phe Tyr 900 905 910

Cys Gly Val Pro Val Leu Gly Lys Glu Leu Ser Lys Leu Cys Asn Thr 915 920 925

Phe Asn Gln Lys Gly Ser Thr Lys Phe Glu Phe His Lys Glu His Phe 930 940

<210> 40

<211> 590

<212> PRT

<213> Rice

<400> 40

Asn Leu Ala Gly Leu Arg Lys Lys Ser Ser Ile Arg Lys Ile Ser Thr 1 5 10 15

- Ser Leu Ser Tyr Tyr Phe Glu Asp Asn Trp Lys Arg Leu Trp Val Leu 20 25 30
- Ala Leu Trp Ile Gly Ile Met Ala Gly Leu Phe Thr Trp Lys Phe Met 35 40 45
- Gln Tyr Arg Asn Arg Tyr Val Phe Asp Val Met Gly Tyr Cys Val Thr
 50 55 60
- Thr Ala Lys Gly Ala Ala Glu Thr Leu Lys Leu Asn Met Ala Ile Ile 65 70 75 80
- Leu Leu Pro Val Cys Arg Asn Thr Ile Thr Trp Leu Arg Ser Thr Arg 85 90 95
- Ala Ala Arg Ala Leu Pro Phe Asp Asp Asn Ile Asn Phe His Lys Thr
- Ile Ala Ala Ala Ile Val Val Gly Ile Ile Leu His Ala Gly Asn His
 115 120 125
- Leu Val Cys Asp Phe Pro Arg Leu Ile Lys Ser Ser Asp Glu Lys Tyr 130 135 140
- Ala Pro Leu Gly Gln Tyr Phe Gly Glu Ile Lys Pro Thr Tyr Phe Thr 145 150 155 160
- Leu Val Lys Gly Val Glu Gly Ile Thr Gly Val Ile Met Val Val Cys
 165 170 175
- Met Ile Ile Ala Phe Thr Leu Ala Thr Arg Trp Phe Arg Arg Ser Leu 180 185 190
- Val Lys Leu Pro Arg Pro Phe Asp Lys Leu Thr Gly Phe Asn Ala Phe
 195 200 205
- Trp Tyr Ser His His Leu Phe Ile Ile Val Tyr Ile Ala Leu Ile Val 210 215 220
- His Gly Glu Cys Leu Tyr Leu Ile His Val Trp Tyr Arg Arg Thr Thr
 225 230 235 240
- Trp Met Tyr Leu Ser Val Pro Val Cys Leu Tyr Val Gly Glu Arg Ile 245 250 255

Leu Arg Phe Phe Arg Ser Gly Ser Tyr Ser Val Arg Leu Leu Lys Val 260 265 270

- Ala Ile Tyr Pro Gly Asn Val Leu Thr Leu Gln Met Ser Lys Pro Pro 275 280 285
- Thr Phe Arg Tyr Lys Ser Gly Gln Tyr Met Phe Val Gln Cys Pro Ala 290 295 300
- Val Ser Pro Phe Glu Trp His Pro Phe Ser Ile Thr Ser Ala Pro Gly 305 310 315 320
- Asp Asp Tyr Leu Ser Ile His Val Arg Gln Leu Gly Asp Trp Thr Arg 325 330 335
- Glu Leu Lys Arg Val Phe Ala Ala Cys Glu Pro Pro Ala Gly Gly 340 345 350
- Lys Ser Gly Leu Leu Arg Ala Asp Glu Thr Thr Lys Lys Ile Leu Pro 355 360 365
- Lys Leu Leu Ile Asp Gly Pro Tyr Gly Ser Pro Ala Gln Asp Tyr Ser 370 380
- Lys Tyr Asp Val Leu Leu Val Gly Leu Gly Ile Gly Ala Thr Pro 385 390 395 400
- Phe Ile Ser Ile Leu Lys Asp Leu Leu Asn Asn Ile Ile Lys Met Glu
 405 410 415
- Glu Glu Glu Asp Ala Ser Thr Asp Leu Tyr Pro Pro Met Gly Arg Asn 420 425 430
- Asn Pro His Val Asp Leu Gly Thr Leu Met Thr Ile Thr Ser Arg Pro 435 440 445
- Lys Lys Ile Leu Lys Thr Thr Asn Ala Tyr Phe Tyr Trp Val Thr Arg
 450 455 460
- Glu Gln Gly Ser Phe Asp Trp Phe Lys Gly Val Met Asn Glu Ile Ala 465 470 475 480
- Asp Leu Asp Gln Arg Asn Ile Ile Glu Met His Asn Tyr Leu Thr Ser
- Val Tyr Glu Glu Gly Asp Ala Arg Ser Ala Leu Ile Thr Met Leu Gln
 500 505 510

Ala Leu Asn His Ala Lys Asn Gly Val Asp Ile Val Ser Gly Thr Lys 515 520 525

Val Arg Thr His Phe Ala Arg Pro Asn Trp Arg Lys Val Leu Ser Lys 530 540

Ile Ser Ser Lys His Pro Tyr Ala Lys Ile Gly Val Phe Tyr Cys Gly 545 550 555 560

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Gly Lys Cys Thr Thr Lys Phe Asp Phe His Lys Glu His Phe 580 585 590

<210> 41

<211> 2619

<212> DNA

<213> Rat

<400> 41

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acagatter agenteracy tegetteracy caactgetat coefficient aggregate 240
tettegaata tettaagtgaa gagaacatte catageatte graetette tegaaggage 300
caccagacag actgeetteg coefficient agenteracy tegetgeetga attetaacag 360
categoryate etgatecet tegetgeaa teetgetee teetgaggg geacetgete 420
attettgaata teetgate gaaageeatt ggateacaac eteacettee ataageetget 480
ggeatatatag acception agenteracy teatacaacy ceacacettee ataageetget 480
acgeracage agaageeaac aggeeatgga tegateetet geeteetgete teetecageet 600
attecateec gagaaagaag atteetgget aaateecate cagteecaa acgtgacagt 660
gatgtatgea geatttacea gtattgetgg cettactgga gtggtegeea etgtggettt 720
ggtteteatg gtaactteag etattggatt tateeggag aattatttg agetetteeg 780

gtatacacat caccttttca tcatctatat catctgctta gggatccatg gcctgggggg 840 gattgtccgg ggtcaaacag aagagagcat gagtgaaagt catccccgca actgttcata 900 ctctttccac gagtgggata agtatgaaag gagttgcagg agtcctcatt ttgtggggca 960 accccctgag tcttggaagt ggatcctcgc gccgattgct ttttatatct ttgaaaggat 1020 ccttcgcttt tatcgctccc ggcagaaggt cgtgattacc aaggttgtca tgcacccatg 1080 taaagttttg gaattgcaga tgaggaagcg gggctttact atgggaatag gacagtatat 1140 attogtamat tgcccctcgm tttccttcct ggmatggcat ccctttactc tgacctctgc 1200 tecagaggaa gaatttttet eeatteatat tegageagea ggggaetgga cagaaaatet 1260 cataaggaca tttgaacaac agcactcacc aatgcccagg atcgaggtgg atggtccctt 1320 tggcacagtc agtgaggatg tcttccagta cgaagtggct gtactggttg gggcagggat 1380 tggcgtcact ccctttgctt ccttcttgaa atctatctgg tacaaattcc agcgtgcaca 1440 caacaagctg aaaacacaaa agatctattt ctactggatt tgtagagaga cgggtgcctt 1500 tgcctggttc aacaacttat tgaattccct ggaacaagag atggacgaat taggcaaacc 1560 ggatttccta aactaccgac tcttcctcac tggctgggat agcaacattg ctggtcatgc 1620 agcattaaac tttgacagag ccactgacgt cctgacaggt ctgaaacaga aaacctcctt 1680 tgggagacca atgtgggaca atgagttttc tagaatagct actgcccacc ccaagtctgt 1740 ggtgggggtt ttcttatgcg gccctccgac tttggcaaaa agcctgcgca aatgctgtcg 1800 geggtaetea agtetggate etaggaaggt teaattetae tteaacaaag aaaegttetg 1860 aattggagga agccgcacag tagtacttct ccatcttcct tttcactaac gtgtgggtca 1920 gctactagat agtccgttgt cgcacaagga cttcactccc atcttaaagt tgactcaact 1980 ccatcattct tgggctttgg caacatgaga gctgcataac tcacaattgc aaaacacatg 2040 aattattatt ggggggattg taaatccttc tgggaaacct gcctttagct gaatcttgct 2100 ggttgacact tgcacaattt aacctcaggt gtcttggttg atacctgata atcttccctc 2160 ccacctgtcc ctcacagaag atttctaagt agggtgattt taaaatattt attgaatcca 2220

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<210> 42

<211> 499

<212> PRT

<213> Rat

<400> 42

Met Val Ile Leu Ile Pro Val Cys Arg Asn Leu Leu Ser Phe Leu Arg

1 5 10 15

Gly Thr Cys Ser Phe Cys Asn His Thr Leu Arg Lys Pro Leu Asp His 20 25 30 .

Asn Leu Thr Phe His Lys Leu Val Ala Tyr Met Ile Cys Ile Phe Thr 35 40 45

Ala Ile His Ile Ile Ala His Leu Phe Asn Phe Glu Arg Tyr Ser Arg
50 55 60

Ser Gln Gln Ala Met Asp Gly Ser Leu Ala Ser Val Leu Ser Ser Leu 65 70 75 80

Phe His Pro Glu Lys Glu Asp Ser Trp Leu Asn Pro Ile Gln Ser Pro 85 90 95

Asn Val Thr Val Met Tyr Ala Ala Phe Thr Ser Ile Ala Gly Leu Thr

Gly Val Val Ala Thr Val Ala Leu Val Leu Met Val Thr Ser Ala Met
115 120 125

Glu Phe Ile Arg Arg Asn Tyr Phe Glu Leu Phe Trp Tyr Thr His His 130 135 140

Leu Phe Ile Ile Tyr Ile Ile Cys Leu Gly Ile His Gly Leu Gly Gly 145 150 155 160

- Ile Val Arg Gly Gln Thr Glu Glu Ser Met Ser Glu Ser His Pro Arg 165 170 175
- Asn Cys Ser Tyr Ser Phe His Glu Trp Asp Lys Tyr Glu Arg Ser Cys 180 185 190
- Arg Ser Pro His Phe Val Gly Gln Pro Pro Glu Ser Trp Lys Trp Ile 195 200 205
- Leu Ala Pro Ile Ala Phe Tyr Ile Phe Glu Arg Ile Leu Arg Phe Tyr 210 215 220
- Arg Ser Arg Gln Lys Val Val Ile Thr Lys Val Val Met His Pro Cys 225 230 235 240
- Lys Val Leu Glu Leu Gln Met Arg Lys Arg Gly Phe Thr Met Gly Ile 245 250 255
- Gly Gln Tyr Ile Phe Val Asn Cys Pro Ser Ile Ser Phe Leu Glu Trp 260 265 270
- His Pro Phe Thr Leu Thr Ser Ala Pro Glu Glu Glu Phe Phe Ser Ile 275 280 285
- His Ile Arg Ala Ala Gly Asp Trp Thr Glu Asn Leu Ile Arg Thr Phe 290 295 300
- Glu Gln Gln His Ser Pro Met Pro Arg Ile Glu Val Asp Gly Pro Phe 305 310 315 320
- Gly Thr Val Ser Glu Asp Val Phe Gln Tyr Glu Val Ala Val Leu Val 325 330 335
- Gly Ala Gly Ile Gly Val Thr Pro Phe Ala Ser Phe Leu Lys Ser Ile 340 345 350
- Trp Tyr Lys Phe Gln Arg Ala His Asn Lys Leu Lys Thr Gln Lys Ile 355 360 365
- Tyr Phe Tyr Trp Ile Cys Arg Glu Thr Gly Ala Phe Ala Trp Phe Asn 370 375 380
- Asn Leu Leu Asn Ser Leu Glu Gln Glu Met Asp Glu Leu Gly Lys Pro 385 390 395 400

Asp Phe Leu Asn Tyr Arg Leu Phe Leu Thr Gly Trp Asp Ser Asn Ile
405 410 415

Ala Gly His Ala Ala Leu Asn Phe Asp Arg Ala Thr Asp Val Leu Thr 420 425 430

Gly Leu Lys Gln Lys Thr Ser Phe Gly Arg Pro Met Trp Asp Asn Glu 435 440 445

Phe Ser Arg Ile Ala Thr Ala His Pro Lys Ser Val Val Gly Val Phe 450 455 460

Leu Cys Gly Pro Pro Thr Leu Ala Lys Ser Leu Arg Lys Cys Cys Arg 465 470 475 480

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Glu Thr Phe

<210> 43

<211> 35

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic primer

<400> 43

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<210> 44

<211> 45

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic
 primer

<400> 44

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45

<210> 45 <211> 5508 <212> DNA <213> Homo sapiens <220> <221> CDS <222> (155)..(4810) <400> 45

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240:	l gctgatgaga	GCAGCTGTGA	CACGGGAGCA	GCGGAGGCAG	CTCCTGGAGA	CCTTTTTCAG
246.	L GCACCTTTTC	: TCCCAGGTGC	: TGGACATCAA	CCAGGCCGAC	GCAGGGACCC	TECCCCTCC
252:	L CTCCTCCCAG	AAGGTGCGGG	AGGCCCTGAC	CTGTGAGCTC	AGCAGGGCCG	AGTTTCCCCA
258:	L GTCCCTGGGC	CTCAAGCCCC	: AGGACATGTT	TGTGGAGTCC	ATGTTCTCTC	TGGCTGACAA
264:	l ggatggcaat	GGCTACCTGT	CCTTCCGAGA	GTTCCTGGAC	ATCCTGGTGG	TCTTCATGAA
2703	LAGGCTCTCCT	' GAGGAAAAGI	CTCGCCTTAT	GTTCCGCATC	TACGACTTTG	ATGGGAATGG
2761	CCTCATTTCC	AAGGATGAGT	'TCATCAGGAT	GCTGAGATCC	TTCATCGAGA	TCTCCAACAA
2821	L CTGCCTGTCC	AAGGCCCAGC	TGGCTGAGGT	GGTGGAGTCC	ATGTTCCGGG	. AGTCGGGATT
2881	CCAGGACAAG	GAGGAACTGA	CATGGGAAGA	TTTTCACTTC	ATGCTGCGGG	ACCACAATAG
2941	CGAGCTCCGC	TTCACGCAGC	TCTGTGTCAA	AGGGGTGGAG	GTGCCTGAAG	TCATCAACCA
3001	CCTCTGCCGG	CGAGCCTCCT	ACATCAGCCA	GGATATGATO	TGTCCCTGTAG	CCDCDCTCAC
3061	TGCCCGCTGT	TCCCGCAGCG	ACATTGAGAC	TGAGTTGACA	CCTCAGAGAC	TCCAGAGIGAG
3121	CATGGACACA	GACCCTCCCC	AGGAGATTCG	GCGGAGGTTT	GGCAAGAAGG	TAACCTCACC
3181	CCAGCCCTTG	CTGTTCACTG	AGGCGCACCG	AGAGAAGTTC	CAACGCAGCT	CTCTCCACCA
3241	GACGGTGCAA	CAGTTCAAGC	GCTTCATTGA	GAACTACCGG	CCCCACATCC	GICICCACCA
3301	CGTGTTCTAC	GCCATCGCTG	GGGGGCTTTT	CCTGGAGAGG	GCCTACTACT	actecetee.
3361	CGCACATCAC	ACGGGCATCA	CGGACACCAC	CCGCGTGGGA	ATCATCCTCT	CCCCCCTCC
3421	AGCAGCCAGC	ATCTCTTTCA	TGTTCTCCTA	CATCTTGCTC	ACCATGTGCC	CCA A CCTCA TO
3481	CACCTTCCTG	CGAGAAACCT	TCCTCAACCG	CTACGTGCCC	TTCGACGCCG	CCCTCCACT
3541	CCATCGCCTC	ATTGCCTCCA	CCGCCATCGT	CCTCACAGTC	TTACACAGTG	TECCCCATCE
3601	GGTGAATGTG	TACCTGTTCT	CCATCAGCCC	CCTCAGCGTC	CTCTCTCTCCC	TOGGCCATGT
3661	CCTCTTCCAT	GATGATGGGT	CTGAGTTCCC	CCAGAAGTAT	TACTGGTGGT	TCTTTCCTGG
3721	CGTACCAGGC	CTCACGGGGG	TTGTGCTGCT	CCTGATCCTG	GCCATCATGT	ATGTCTTTTCC
3781	CTCCCACCAC	TTCCGCCGCC	GCAGTTTCCG	GGGCTTCTGG	CTGACCCACC	ACCTCTACAT
3841	CCTGCTCTAT	GTCCTGCTCA	TCATCCATGG	TAGCTTTGCC	CTGATCCAGC	TECCCCCTT
3901	CCACATCTTC	TTCCTGGTCC	CAGCAATCAT	CTATGGGGGC	GACAAGCTGG	TGAGCCTGAC
3961	CCGGAAGAAG	GTGGAGATCA	GCGTGGTGAA	GGCGGAGCTG	CTGCCCTCAG	GAGTCACCCA
4021	CCTGCGGTTC	CAGCGGCCCC	AGGGCTTTGA	GTACAAGTCA	GGGCAGTGGG	TECEGATECE
4081	TTGCCTGGCT	CTGGGGACCA	CCGAGTACCA	CCCCTTCACA	CTGACCTCTG	CCCCCCATCA
4141	GGACACGCTT	AGCCTGCACA	TCCGGGCAGC	AGGGCCCTGG	ACCACTCGCC	TCAGGGAGAT
4201	CTACTCAGCC	CCGACGGGTG	ACAGATGTGC	CAGATACCCA	AAGCTGTACC	TEATERICACE
4261	ATTTGGAGAG	GGCCACCAGG	AGTGGCATAA	GTTTGAGGTG	TCAGTGTTAG	TEGGIEGEG
4321	CATTGGGGTC	ACCCCTTTTG	CCTCCATCCT	CAAAGACCTG	GTCTTCAAGT	CATCCCTCAC
4381	CTGCCAAGTG	TTCTGTAAGA	AGATCTACTT	CATCTGGGTG	ACGCGGACCC	ACCCACACAM
4441	TGAGTGGCTG	GCTGACATCA	TCCGAGAGGT	GGAGGAGAAT	GACCACCAGG	ACCTCCTCTC
4501	TGTGCACATC	TACATCACCC	AGCTGGCTGA	GAAGTTCGAC	CTCAGGACCA	CTATCCTCTA
4561	CATCTGTGAG	CGGCACTTCC	AGAAGGTTCT	GAACCGGAGT	CTATTCACAG	CIAIGCIGIA
4621	CATCACCCAC	TTTGGCCGTC	CCCCCTTTGA	GCCCTTCTTC	AACTCCCTCC	ACCACCTOCA
4681	CCCCCAGGTC	CGGAAGATCG	GGGTGTTTAG	CTGTGGCCCC	CCTGGCATGA	CCFFCF FACE ALCOHOLD
4741	GGAAAAGGCC	TGTCAGCTCA	TCAACAGGCA	GGACCGGACT	CACTTOTOCC	DCCDAMANCA
4801	GAACTTCTAG	GCCCCTGCCC	GGGGGTTCTG	CCCACTGTCC	AGTTGAGCAG	ACCRITATION
4861	CCACACCTCA	CCTCTGTTCT	TCCTATTTCT	GGCTGCCTCA	GCCTTCTCTC	ATTICCCACC
4921	TCCCAACCTT	GTTCCAGGTG	GCCATAGTCA	GTCACCATGT	GTGGGCTCAG	GGACCCCCAC
4981	GACCAGGATG	TGTCTCAGCC	TGGAGAAATG	GTGGGGGGC	AGTGTCTAGG	GACTAGACTC
5041	AGAAGTAGGG	GAGCTACTGA	TTTGGGGCAA	AGTGAAACCT	CTGCTTCAGA	CTTCAGAAAC
2101	AAATCTCAGA	AGACAAGCTG	ACCTGACAAG	TACTATGTGT	GTGCATGTCT	GTATGTGTGT
2161	TGGGGCGGTG	AGTGTAAGGA	TGCAGTGGGA	GCATGGATGC	TGGCATCTTA	GAACCCTCCC
5221	TACTCCCATA	CCTCCTCCTC	TTCTGGGCTC	CCCACTGTCA	GACGGGCTGG	СУУЛЕСССТ
5281	GCAGGAGGTA	GAGGCTGGAC	CCATGGCAAG	CCATTTACAG	AAACCCACTC	GGCACCCCAC
		•		· -		

5341 TCTAACACCA CAACTAATTT CACCCAAGGT TTTAAGCACG TTCTTTCATC AGACCCTGGC

5401 CCAATACCTA TGTATGCAAT GCTCCTCAGC CCTCTTCTCC CTGCTCCAGT AGTCTCCCTT

5461 ССАААТАААТ САСТТТТСТС ССААААААА АААА

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<212> PRT

<213> Homo sapiens

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121	EFLNIRIPPG	DPMFDPDQRG	DVVLPFQRSR	WDPETGRSPS	NPRDPANOVT	GWLDGSATVG
181	SSHSWSDALR	SFSRGQLASG	PDPAFPRDSQ	NPLLMWAAPD	PATGONGPRG	LYAFGAERGN
241	REPFLQALGL	LWFRYHNLWA	QRLARQHPDW	EDEELFOHAR	KRVIATYONT	AVVEWI.DOET.
301	QKTLPEYTGY	RPFLDPSISS	EFVAASEQFL	STMVPPGVYM	RNASCHFOGV	TNRNSSVSRA
361	LRVCNSYWSR	EHPSLQSAED	VDALLLGMAS	QIAEREDHVL	VEDVRDFWPG	PLKESPTDHI.
421	ASCLQRGRDL	GLPSYTKARA	ALGLSPITRW	QDINPALSRS	NDTVLEATAA	LYNODI,SWI.F
481	LLPGGLLESH	RDPGPLFSTI	VLEQFVRLRD	GDRYWFENTR	NGLESKKETE	ETRNTTTI ODU
541	LVAVINIDPS	ALQPNVFVWH	KGDPCPQPRQ	LSTEGLPACA	PSVVRDYFEG	SGEGEGUTTC
601	TLCCFPLVSL	LSAWIVARLR	MRNFKRLQGQ	DROSIVSEKL	VGGMEALEWO	GHKEPCRDIA.
661	VYLQPGQIRV	VDGRLTVLRT	IQLQPPQKVN	FVLSSNRGRR	TLLLKIPKEY	DIATA ENLER
721	ERQALVENLR	GALKESGLSI	QEWELREQEL	MRAAVTREOR	RHLLETFFRH	LESOVEDINO
781	ADAGTLPLDS	SQKVREALTC	ELSRAEFAES	LGLKPODMFV	ESMFSLADKD	CNCVI.SEREE
841	LDILVVFMKG	SPEEKSRLMF	RMYDFDGNGL	ISKDEFIRML	RSFIEISNNC	LSKAOLAFAN
901	ESMFRESGFQ	DKEELTWEDF	HFMLRDHNSE	LRFTOLCVKG	VEVPEVIKDI.	CEPACVICON
961	MICPSPRVSA	RCSRSDIETE	LTPQRLQCPM	DTDPPQEIRR	RFGKKVTSFO	PLI.FTEAUDE
1021	KFQRSCLHQT	VQQFKRFIEN	YRRHIGCVAV	FYAIAGGLFL	ERAYYYAFAA	HHTCTTIVTTD
1081	VGIILSRGTA	ASISFMFSYI	LLTMCRNLIT	FLRETFLNRY	VPFDAAVDEH	PI.TA CTATUT
1141	TVLHSVGHVV	NVYLFSISPL	SVLSCLFPGL	FHDDGSEFPO	KYYWWFFOTV	PGI/TGVVI.I.I.
1201	ILAIMYVFAS	HHFRRRSFRG	FWLTHHLYIL	LYVLLIIHGS	FALTOLPREH	TEELUDATTY
1261	GGDKLVSLSR	KKVEISVVKA	ELLPSGVTHL	RFORPOGFEY	KSGOWVRTAC	LALCTTEVED
1321	FTLTSAPHED	TLSLHIRAAG	PWTTRLREIY	SAPTGDRCAR	YPKLYLDGPF	GEGRUERINGE
1381	EVSVLVGGGI	GVTPFASILK	DLVFKSSVSC	QVFCKKIYFI	WVTRTOROFE	WI.ADTTREVE
1441	ENDHQDLVSV	HIYITQLAEK	FDLRTTMLYI	CERHFOKVLN	RSLFTGLRST	THECRPPER
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<211> 3453

<212> DNA

<213> Homo sapiens

<220>

<221> CDS

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<400> 47

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tggtcgctgt tatcaacatt gaccccagtg ccctgcagcc caatgtcttt gtctggcata 180

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ecctgactgt gettgactte tttgaaggea geagecetgg ttttgecate accateattg 300

ctctctgctg ccttccctta gtgagtctgc ttctctctgg agtggtggcc tatttccggg 360

geegagaaca caagaageta caaaagaaac teaaagagag egtgaagaag gaageageea 420

aagatggagt gccagcg atg gag tgg cca ggc ccc aag gag agg agc agt 470

Met Glu Trp Pro Gly Pro Lys Glu Arg ser ser

1 5 10

ccc atc atc atc cag ctg ctg tca gac agg tgt ctg cag gtc ctg aac 518
Pro Ile Ile Ile Gln Leu Leu Ser Asp Arg Cys Leu Gln Val Leu Asn
15 20 25

agg cat ctc act gtg ctc cgt gtg gtc cag ctg cag cct ctg cag cag 566
Arg His Leu Thr Val Leu Arg Val Val Gln Leu Gln Pro Leu Gln Gln
30 35 40

gtc aac ctc atc ctg tcc aac cga gga tgc cgc acc ctg ctg ctc 614
Val Asn Leu Ile Leu Ser Asn Asn Arg Gly Cys Arg Thr Leu Leu Leu
45 50 55

aag atc cct aag gag tat gac ctg gtg ctg ctg ttt agt tct gaa gag
Lys Ile Pro Lys Glu Tyr Asp Leu Val Leu Leu Phe Ser Ser Glu Glu
60 75

gaa cgg ggc gcc ttt gtg cag cag cta tgg gac ttc tgc gtg cgc tgg 710 Glu Arg Gly Ala Phe Val Gln Gln Leu Trp Asp Phe Cys Val Arg Trp 80 85 90

gct ctg ggc ctc cat gtg gct gag atg agc gag aag gag cta ttt agg 758

		, 0 00														PC
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aag Lys	gct Ala	gtg Val 110	aca Thr	aag Lys	cag Gln	cag Gln	cgg Arg 115	gaa Glu	cgc Arg	atc Ile	ctg Leu	gag Glu 120	atc Ile	ttc Phe	ttc Phe	806
aga Arg	cac His 125	ctt Leu	ttt Phe	gct Ala	cag Gln	gtg Val 130	ctg Leu	gac Asp	atc Ile	aac Asn	cag Gln 135	gcc Ala	gac Asp	gca Ala	ej aaa	854
acc Thr 140	ctg Leu	ccc Pro	ctg Leu	gac Asp	tcc Ser 145	tcc Ser	cag Gln	aag Lys	gtg Val	cgg Arg 150	gag Glu	gcc Ala	ctg Leu	acc Thr	tgc Cys 155	902
gag Glu	ctg Leu	agc Ser	agg Arg	gcc Ala 160	gag Glu	ttt Phe	gcc Ala	gag Glu	tcc Ser 165	ctg Leu	ggc	ctc Leu	aag Lys	ccc Pro 170	cag Gln	950
gac Asp	atg Met	ttt Phe	gtg Val 175	gag Glu	tcc Ser	atg Met	ttc Phe	tct Ser 180	ctg Leu	gct Ala	gac Asp	aag Lys	gat Asp 185	GJ Y ggc	aat Asn	998
Gly ggc	tac Tyr	ctg Leu 190	tcc Ser	ttc Phe	cga Arg	gag Glu	ttc Phe 195	ctg Leu	gac Asp	atc Ile	ctg Leu	gtg Val 200	gtc Val	ttc Phe	atg Met	1046
							Ser	cgt Arg								1094
								aag Lys								1142
								aac Asn					-	-	_	1190
gcc Ala	gag Glu	gtg Val	gtg Val 255	gag Glu	tct Ser	atg Met	ttc Phe	cgg Arg 260	gag Glu	tcg Ser	gga Gly	ttc Phe	cag Gln 265	gac Asp	aag Lys	1238
gag Glu	gag Glu	Leu 270	aca Thr	tgg Trp	gag Glu	gat Asp	ttt Phe 275	cac His	ttc Phe	atg Met	ctg Leu	cgg Arg 280	gac Asp	cat His	gac Asp	1286
agc	gag	ctc	cgc	ttc	acg	cag	ctc	tgt	gtc	aaa	ggt	gga	ggt	gga	ggt	1334

52

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Ser	Glu 285	Leu	Arg	Phe	Thr	Gln 290	Leu	Cys	Val	Lys	Gly 295	Gly	Gly	Gly	Gly	
														cga Arg		1382
														gga Gly 330		1430
														ctg Leu		1478
														tac Tyr		1526
														aag Lys		1574
cag Gln 380	cag Gln	tac Tyr	aag Lys	cgc Arg	ttc Phe 385	gtg Val	gag Glu	aac Asn	tac Tyr	cgg Arg 390	agg Arg	cac His	atc Ile	gtg Val	tgt Cys 395	1622
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														acc Thr		1718
														tcc Ser		1766
														acc Thr		1814
														gca Ala		1862
gac	ttc	cac	cgc	tgg	atc	gcc	atg	gct	gct	gtt	gtc	ctg	gcc	att	ttg	1910

53

				_												PCI
Asp	Phe	His	Arg	Trp 480	Ile	Ala	Met	Ala	Ala 485	Val	Val	Leu	Ala	Ile 490		
cac	agt	act	aac	cac	σca	atc	aat	gtc	tac	a t c	++-	•				
His	Ser	Ala	Glv	His	Ala	Val	Den	Val	Tur	Tio	Db	tca	gtc	agc	cca	1958
			495		744	,,,	7311	500	IYL	116	Pne	Ser	Val 505	Ser	Pro	
ctc	agc	ctg	ctg	gcc	tgc	ata	ttc	ccc	aac	gtc	ttt	ata	aat	gat	aaa	2006
Leu	Ser	Leu	Leu	Ala	Cys	Ile	Phe	Pro	Asn	Val	Phe	Val	Asn	Asp	Glv	2000
		510					515					520		,	or,	
tcc	aag	ctt	ccc	cag	aag	ttc	tat	tgg	tgg	ttc	ttc	cag	acc	gtc	cca	2054
Ser	Lys	Leu	Pro	Gln	Lys	Phe	Tyr	Trp	Trp	Phe	Phe	Gln	Thr	Val	Pro	
	525					530					535					
ggt	atg	aca	ggt	gtg	ctt	ctg	ctc	ctg	gtc	ctg	gcc	atc	atg	tat	gtc	2102
Gly	Met	Thr	Gly	Val	Leu	Leu	Leu	Leu	Val	Leu	Ala	Ile	Met	Tyr	Val	
540					545					550					555	
ttc	gcc	tcc	cac	cac	ttc	cgc	cgc	cgc	agc	ttc	cgg	ggc	ttc	tgg	ctg	2150.
Phe	Ala	Ser	His		Phe	Arg	Arg	Arg	Ser	Phe	Arg	Gly	Phe	Trp	Leu	
				560					565					570		
acc	cac	cac	ctc	tac	atc	ctg	ctc	tat	gcc	ctg	ctc	atc	atc	cat	ggc	2198
Thr	His	His		Tyr	Ile	Leu	Leu	Tyr	Ala	Leu	Leu	Ile	Ile.	His	Gly	
			575					580					585			
agc	tat	gct	ctg	atc	cag	ctg	ccc	act	ttc	cac	atc	tac	ttc	ctg	gtc	2246
Ser	Tyr		Leu	Ile	Gln	Leu	Pro	Thr	Phe	His	Ile	Tyr	Phe	Leu	Val	
		590					·595					600				
ccg	gca	atc	atc	tat	gga	ggt	gac	aag	ctg	gtg	agc	ctg	agc	cgg	aag	2294
Pro	Ala	Ile	Ile	Tyr	Gly		Asp	Lys	Leu	Val	Ser	Leu	Ser	Arg	Lys	
	605					610					615					
aag	gtg	gag	atc	agc	gtg	gtg	aag	gcg	gag	ctg	ctg	CCC	tca	gga	gtg	2342
Lys	Val	Glu	Ile	Ser		Val	Lys	Ala	Glu	Leu	Leu	Pro	Ser	Gly	Val	
620					625				•	630					635	
acc	tac	ctg	caa	ttc	cag	agg	ccc	caa	ggc	ttt	gag	tac	aag	tca	gga	2390
Thr	Tyr	Leu	Gln	Phe	Gln	Arg	Pro	Gln	Gly	Phe	Glu	Tyr	Lys	Ser	Gly	
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Ile	Arg	Ala	Val	999 Glv	Pro	Tro	Thr	Thr	Ara	T.e.	agg	gag	ato	tac	tca	2534
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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification 7:

C12N 15/12, 15/53, C07K 16/40, 16/18, 14/47, C12N 9/02, A61K 38/17, 38/44, 48/00, G01N 33/50

(11) International Publication Number:

WO 00/28031

(43) International Publication Date:

18 May 2000 (18.05.00)

(21) International Application Number:

PCT/US99/26592

A2

(22) International Filing Date:

10 November 1999 (10.11.99)

(30) Priority Data:

60/107,911 10 November 1998 (10.11.98) US 60/149,332 17 August 1999 (17.08.99) US 60/151,242 27 August 1999 (27.08.99) US

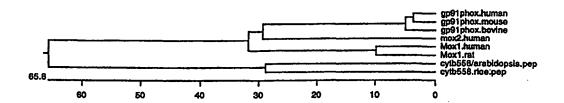
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- (72) Inventors: LAMBETH, J., David; 461 Emory Drive, Decatur, GA 30207 (US). LASSEGUE, Bernard, P.; 564 Emory Oaks Way, Decatur, GA 30033 (US). GRIENDLING, Kathy, K.; 1819 Kanawha Trail, Stone Mountain, GA 30087 (US). ARNOLD, Rebecca, S.; 4344 Bridle Path Court, Tucker, GA 30084 (US). GUANGJIE, Cheng; Apt 1, 2751 Briarcliff Road, Atlanta, GA 30329 (US).
- (74) Agents: ELSEVIER, Lisa, C. et al.; Jones & Askew, LLP, 2400 Monarch Tower, 3424 Peachtree Road, N.E., Atlanta, GA 30326 (US).

(81) Designated States: AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK, DM, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, UZ, VN, YU, ZA, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

Published

Without international search report and to be republished upon receipt of that report.

(54) Title: NOVEL MITOGENIC REGULATORS



(57) Abstract

The present invention relates to new genes encoding for the production of novel proteins involved in generation of reactive oxygen intermediates that affect cell division. The present invention also provides vectors containing these genes, cells transfected with these vectors, antibodies raised against these novel proteins, kits for detection, localization and measurement of these genes and proteins, and methods to determine the activity of drugs to affect the activity of the proteins of the present invention.

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(43) International Publication Date:

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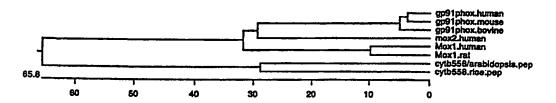
Published

With international search report.

(88) Date of publication of the international search report:

23 November 2000 (23.11.00)

(54) Title: MITOGENIC REGULATORS



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PCT/US 99/26592 A. CLASSIFICATION OF SUBJECT MATTER IPC 7 C12N15/12 C12N C12N15/53 C07K16/40 C07K14/47 C07K16/18 C12N9/02 A61K38/17 A61K38/44 A61K48/00 G01N33/50 According to International Patent Classification (IPC) or to both national classification and IPC **B. FIELDS SEARCHED** Minimum documentation searched (classification system followed by classification symbols) IPC 7 C12N C07K A61K G01N Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Electronic data base consulted during the international search (name of data base and, where practical, search terms used) EPO-Internal, STRAND C. DOCUMENTS CONSIDERED TO BE RELEVANT Category 5 Citation of document, with indication, where appropriate, of the relevant passages Relevant to claim No. X LI F ET AL: "CD34+ PERIPHERAL BLOOD 1,3,5,7, PROGENITORS AS A TARGET FOR GENETIC 9,11,13 CORRECTION OF THE TWO FLAVOCYTOCHROME B558 DEFECTIVE FORMS OF CHRONIC GRANULOMATOUS DISEASE" BLOOD, US, W.B. SAUNDERS, PHILADELPHIA, VA, vol. 84, no. 1, 1 July 1994 (1994-07-01), pages 53-58, XP000674233 ISSN: 0006-4971 A page 54, column 2 2,4.6.8. 10,12,14 figures 1,2 X US 5 593 966 A (MALECH HARRY L ET AL) 1,3,9,15 14 January 1997 (1997-01-14) Α column 5, line 34 -column 6, line 53; 2,4,10, figure 2; table 1 -/---Further documents are listed in the continuation of box C. Patent family members are listed in annex. * Special categories of cited documents : "I later document published after the international filing date or priority date and not in conflict with the application but "A" document defining the general state of the art which is not considered to be of particular relevance cited to understand the principle or theory underlying the invention "E" earlier document but published on or after the international "X" document of particular relevance; the claimed invention filing date cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled "O" document referring to an oral disclosure, use, exhibition or other means document published prior to the international filing date but later than the priority date claimed "&" document member of the same patent family Date of the actual completion of the international search Date of mailing of the international search report 31.08.00 16 August 2000 Name and mailing address of the ISA Authorized officer European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+31-70) 340-3016

van Klompenburg, W

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C.(Continu	Citation of document, with indication where appropriate, of the relevant passages	
	order of document, with inducation, where appropriate, of the relevant passages	Relevant to daim No.
X	STRAUSBERG ET AL.: "National cancer institute, cancer genome anatomy project (CGAP)" EMBL DATABASE ACC NO: AA493362, 28 June 1997 (1997-06-28), XP002137597 the whole document	1-14
X	LLOYD: "Human DNA sequence from clone 146h21 on chromosome Xq22" EMBL DATABASE ACC NO: Z83819, 10 January 1997 (1997-01-10), XP002137598 the whole document	1-14
X	ADAMS ET AL.: "Initial assessment of human gene diversity and expression patterns based upon 83 million basepairs of cDNA sequence" EMBL DATABASE ACC NO: AA305700, 18 April 1997 (1997-04-18), XP002137621 cited in the application	1-10
Α	the whole document	11-16
X	PALMER: "Human DNA sequence from clone 25719 on chroosome 6q25.1-26 contains gene similar to Cytochrome B, CA repeat, GSS" EMBL DATABASE ACC. NO.: ALO31773, 29 September 1998 (1998-09-29), XP002144975 the whole document	1-14
X	HILLIER ET AL.: "Generation and analysis of 280,000 human expressed sequence tags" EMBL DATABASE ACC. NO.: W52750, 4 June 1996 (1996-06-04), XP002144976 the whole document	1-10
X	STRAUSBERG: "National cancer institute, cancer genome anatomy project (CGAP)" EMBL DATABASE ACC. NO.: AA641653, 1 November 1997 (1997-11-01), XP002144977 the whole document	1-10
A	WILSON ET AL.: "f53g12.3" EMBL DATABASE ACC. NO.: AF003139, 1 July 1997 (1997-07-01), XP002144978 abstract	1-16
P,X	SUH ET AL.: "Cell transformation by the superoxide-generating oxidase Mox1" NATURE, vol. 401, 2 September 1999 (1999-09-02), pages 79-82, XP002137599 figures 1-5	1-8, 11-14
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	ation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication where appropriate, of the relevant passages		Relevant to claim No.
T	DUPUY ET AL.: "Purification of a novel flavoprotein involved in the thyroid NADPH oxidase" THE JOURNAL OF BIOLOGICAL CHEMISTRY, vol. 274, no. 52, 24 December 1999 (1999–12–24), pages 37265–37269, XP002144979 figures 2,3 -& DUPUY ET AL.: EMBL DATABASE ACC. No.: AF181972, 29 December 1999 (1999–12–29), XP002144980 the whole document		1-8

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Box	Observations where certain claims were found unsearchable (Continuation of Item 1 of first sheet)
This Inte	emational Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
1. X	Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
	In so far as claims 11-14 are directed to a method of treatment of the human/animal body, the search has been carried out and based on the alleged effects of the compound/composition.
2.	Claims Nos.: because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
3.	Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II	Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
This Inte	ernational Searching Authority found multiple inventions in this international application, as follows:
	see additional sheet
1. 🔽	As all required additional search fees were timely paid by the applicant, this International Search Report covers all
نـهــا	searchable claims.
2.	As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3.	As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
	·
4.	No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Remark	k on Protest The additional search fees were accompanied by the applicant's protest.
	No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

This International Searching Authority found multiple (groups of) inventions in this international application, as follows:

1. Claims: 1-16 all partially

A protein capable of stimulating superoxide production, wherein the protein comprises a mox1, preferably with the sequence of SEQ ID NO 2, SEQ ID NO 21, SEQ ID NO 42, a fragment thereof or a conservative substitution thereof. A nucleotide sequence, preferably with the sequence of SEQ ID NO 1, SEQ ID NO 22 or SEQ ID NO 41 encoding for the above mentioned protein, fragment thereof or conservative substitution.

A vector comprising said nucleotide sequence and a cell containing said vector.

An antibody capable of binding the above mentioned protein, fragment or conservative substitution. A method of stimulating superoxide formation, in vitro or in vivo, comprising administration, in vitro or in vivo, of a composition comprising the abovementioned vector or the above mentioned protein or its fragment or its conservative substitution in a pharmaceutically acceptable vector. A method for determining the activity of a drug comprising measuring the activity of the above mentioned protein to stimulate superoxide formation following administration of the drug.

2. Claims: 1-16 all partially

As invention 1, but for a mox2 with SEQ ID NO: 4 and SEQ ID NO: 3 $\,$

3. Claims: 1-16 all partially

As invention 1, but for a duox1 with SEQ ID NO: 46 and SEQ ID NO: 45 and for a duox2 with SEQ ID NO: 48 and SEQ ID NO: 47

Information on patent family members

In. ational Application No
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Patent document cited in search report		Publication date	Patent family member(s)			Publication date
US 5593966	A	14-01-1997	US AU WO	5585346 7956791 9117763	Α	17-12-1996 10-12-1991 28-11-1991

Form PCT/ISA/210 (patent family annex) (July 1992)